



Second Generation Biofuel – An Alternate Source of Energy

R. Devika*, M. Subathra, J. Madhusudhanan, Rajesh and UditSagar

*Department of Biotechnology, AarupadaiVeedu Institute of Technology, Vinayaka Missions Research Foundation (Deemed to be University),
Paiyanoor - 603 104, Tamilnadu, India.*

Abstract: The use of fossil fuels in the current situation has been increased. These fossil fuels do not come under the category of sustainable sources due to its depleting nature. Fossil fuels also pollutes the environment by increased emission of Greenhouse Gases (GHG). In order to produce an alternate renewable energy and to reduce the percentage of environmental pollution caused, many technologies like solar energy, tidal energy, wind energy etc. came into existence. But all these forms of energy can be produced in terms of electricity. To meet the increasing fuel demand, renewable biofuels came into the field where biomasses are utilized to produce biofuels. Second generation renewable biofuels are produced from conventional plant sources with higher oil content whose efficiency is determined by the amount of Green House Gases (GHG) emitted and the life cycle cost whether positive, neutral or negative. High yielding energy crops such as sugarcane, corn, switchgrass, wheat etc. are subjected to various processes / fermentation to yield biofuels like bioethanol, biobutanol, biohydrogen, biodiesel etc. The US and Brazil are replacing the demand for gasoline with around 15% of bioethanol and it has a higher octane rating, increased engine's compression ratio, increased thermal efficiency and ultimately reduces atmospheric pollution emissions. In future biofuels will be recognized as alternative fuel in transportation, energy generation, heat producer, charging electronics, environmental friendly cleaners, cooking, lubricants, adhesives etc. This review provides a brief outline about the second generation biofuels which would serve as an alternative energy in the near future.

Keywords: Biofuel, saccharification, liquefaction, lignocelluloses, cellulose, lignin, fermentation.

*Corresponding Author

Dr.R.Devika, M.Sc.,M.Phil.,PhD.,PGDBI.,M.Tech.,PhD.,,
Professor and Head, Department of Biotechnology,
AarupadaiVeedu Institute of Technology, Paiyanoor- 603 104,
Chennai, Tamilnadu, India.



Received On 20 June 2020

Revised On 30 October 2020

Accepted On 06 November 2020

Published On 04 January 2021

Funding This research did not receive any specific grant from any funding agencies in the public, commercial or not for profit sectors.

Citation R. Devika, M. Subathra, J. Madhusudhanan, Rajesh and UditSagar , Second Generation Biofuel – An Alternate Source of Energy.(2021).Int. J. Life Sci. Pharma Res.11(1), L238-245 <http://dx.doi.org/10.22376/ijpbs/lpr.2021.11.1.L238-245>

This article is under the CC BY- NC-ND Licence (<https://creativecommons.org/licenses/by-nc-nd/4.0/>)



Copyright @ International Journal of Life Science and Pharma Research, available at www.ijlpr.com

I. INTRODUCTION

Biofuel production research in the current scenario has dramatically increased due intensification of civilization, transport, demand for fossil fuels etc.¹. The global demand has been reported as 32.4 billion gallons in 2013 and it is predicted to increase to 51.1 billion gallons in 2022². It has been depicted that about 13 fold energy consumption increased in the 20th century which is greater than the rate of population growth³⁻⁵. Predicted analysis between 2006 and 2013 revealed that 80% of fuel consumption was against transport and it has diminished the supply of fossil fuels and stimulated the use and production of biofuels as alternative source⁶⁻¹⁰. Evidence of biofuel consumption in EU27 countries from 2006 and 2008 was 5625 Kilotons of oil to 10064 Kilotons where the biofuel production increased from 5639 million tons to 8165 million tons¹¹. Anticipated rising energy demand has developed to an alternate method of usage of biomass for energy production^{12,13}. Biomass waste (maize, sugarcane, sugar beets, rice husk) were worldwide used for production of biofuels in most developing countries¹⁴. The energy from biofuel is considered to be a promising sustainable alternative source for its eco friendly, economic nature for future¹⁵. Nowadays sources for production of biofuel are well studied and reviewed that the availability of bioenergy from feedstock had been recommended for its special and abundant availability of cellulosic and lignocellulosic nature^{16,17}. India, is known for its agricultural resources and enormous supply of waste is wasted (1/3rd as waste) every year^{18,19}. Developing countries which depend on natural resources grow energy crops (eg) Brazil which is located mainly between Equator and Tropic of Capricorn is growing crops for about 851 million hectares²⁰. With 30 years of experience, Brazil has developed bioethanol industries from sugarcane (since the climatic conditions suit the growth and high yield)²¹. A study from NIPE/ UNICAMP stated that Brazilian Government encourages the increase of sugarcane production to meet the demand of gasoline in 2025 and this can be overcome by substituting ethanol (5% 102 billion litres)²². Agro industrial traders are acting as cross border in biofuel production in US and European countries followed by Canada and Mexico^{23,24}. Use of maize has shifted towards use of jatropha and palm oil as alternative feed stocks in Central America and Soy in neighboring Uruguay, Paraguay and Bolivia²⁵. The present review article focuses on the importance and strategic plans adopted in various

countries for production of biofuels from biomass (wealth from waste).

I.I CHARACTERISTICS OF BIOFUEL

It has been predicted that by 2050, quarter of the world will depend on biofuel for transport fuel because of low carbon, non petroleum fuel, minimal changes to vehicle stock²⁶ and this will support the economic development for the country²⁷. Biofuel usage has substantially increased from 2012 (25%) of anhydrous and hydrated ethanol (19.1 Mm³) and biodiesel (2.2 Mm³)²⁸. Research on Greenhouse Gas Emission (GMG threshold) reduction by Renewable Fuel Standard (RFS) the biofuel should have 50% lower emission and it will classified as "Advanced Biofuel" and 60% as "Cellulosic Biofuel" and other biomass registered 44% (Corn ethanol) and 68% (sugar cane ethanol)^{29,30}. Experimental analysis of biobutanol with gasoline in various compositions (100% Butanol, 50% gasoline and Butanol and 95% gasoline) were listed in the city of Sao Paulo for 120 km. It was found that the emission of biogenic CO₂ was 231.5 by 100% butanol and 104.2 in the second combination and 9.53 in the third case of 95% gasoline and CO₂ was nil in 100% butanol and 127.3 and 221.09 in other cases³¹. Elementary analysis of biocrude oil revealed carbon (75.2%), hydrogen (8.2%), nitrogen (0.5%), sulphur (0.3%), oxygen (15.8%), ash content (0.48%), bound water (3.8%)^{32,33}. The cellulosic biocrude through Hydrothermal Liquefaction consisted of 4.4 mg/g aldehyde which was present as 5 – hydro-xymethyl-furfural (5-HMF) and 3.9 mg/g of ketone as 3-methyl-1,2 cyclopentanedione, 2,5-hexanedione and 1-(2-furanyl)ethane etc. and phenolic compounds such as guaiacol and creosol³⁴. The leading bioethanol producers are Brazil with 19,000 million litres (ML) which is equivalent to 10.44 million litres of oil (MTOE) followed by China with 1,840 ML with 1.01 MTOE which is followed by Canada of 1,000 ML with 0.55 MTOE and USA ranked first with 26,500 ML and 14.55 MTOE³⁵. Bioethanol comprises of oxygen (35%) which influence complete combustion of fuel and imparts reduced particulate emission^{36,37} and even 10% ethanol blends GHG emission to 12.19% when compared to fossil fuel source³⁸ and 85% ethanol as fuel reduces nitrogen oxide to 10%, particulate to 20% and sulphate emission to 80%, respectively compared to conventional gasoline. The out/input ratio of various biomass is tabulated in Table I.

Table I : Out/input ratio of various biomasses

Crop Biomass	Output / input ratio	References
Corn	6.9 - 9.5	39
Alfalfa corn	2.9-3.1	40
Switchgrass	10.8-11.3	40
Sugar beet	1.8	41
Winter cereals	2.5-2.8	42

I.2 BIOFUEL AS ALTERNATIVE ENERGY SOURCE

Around 16 distilleries in Pakistan are functioning effectively and produce 506.33 Million Litres (ML) of alcohol from 1.687 Million Tonnes of molasses⁴³. It is also reported that Pakistan is exporting ethanol to European Union around 141.3 ML in 2004, 212.16 in 2006 etc^{44,45}. In Brazil 25% of sugarcane production is utilized for 25% butanol and 50% of ethanol production⁴⁷. The second generation biobutanol has more advantages that the photochemical oxidation was reduced

from 30% to 20%⁴⁸ and this has proved that bioethanol has been considered eco friendly fuel in Brazil²⁸. More recently, Brazil started concentrating on corn fermentation with yeast cells and proved that the process was faster (34-36h) when compared to the traditional process (45-60h)^{49,50}. Dry weight of sugarcane bagasse contain 60% of lignocellulosic and cellulose sources which are accountable for ethanol production by acid or enzymatic hydrolysis⁵¹. Researchers still have technological challenges to overcome the pretreatment process, use of suitable enzymes, improve the

efficiency of bagasse hydrolysis etc. for production of highly efficient second generation ethanol and butanol^{52,53}. Hemicellulosic has been classified as micro and cellulose as macro fibrils⁵⁴ and lignin provides the structural role of the matrix⁵⁵ and these are demonstrated to be the major constituents for biofuel production by enzymatic hydrolysis⁵⁶. First generation ethanol was produced basically from sugarcane and corn worldwide and production was 25 billion gallons where Brazil and the US contributed 85% globally⁵⁷. In the second generation of biofuels low valued lignocellulosic materials from forestry, agriculture were used for production wherein improved pretreatment, fermentation processes were subjected to meet the challenges in 20th century⁵⁸. The application of thermostable enzymes was used to achieve ethanol production of 40-50%⁵⁹ assisting liquefaction process. Characterization of bioethanol production were performed enzymatically using many acids of different concentrations (eg. Sulphuric, hydrochloric, hydrofluoric, formic phosphoric and nitric acids)⁶⁰ and with common catalyst⁶¹ and varied temperature⁶². Three best fermentation process for ethanol production from lignocellulosic sources are Separate Hydrolysis and Fermentation (SHF), Saccharification and Fermentation (SF) and Simultaneous Saccharification and fermentation (SSCF) etc^{63,64} and the microbial strains plays an important role for efficient production⁶⁵ and it is proved

Saccharomyces cereviae as ultimate choice of strain⁶⁶ and *Scheffersomyces shehatae* (*Candida shehatae*) is also proved to be promising in ethanol production⁶⁶.

1.3 ADVANTAGES OF SECOND GENERATION BIOFUELS

The biofuels obtained from sugar and vegetables oils are classified under first generation biofuels. Whereas, the second generation biofuels are obtained from cellulose, hemicelluloses or lignin sources. The advantages of second generation biofuels are

- Less greenhouse gas emission (upto 86%)
- Fully biodegradable
- Environmentally friendly
- It can be directly supported or blended with petrol in some proportion (ethanol)
- Lower energy density
- Enhance and safeguard energy security⁶⁷

The second generation technology includes processes like thermochemical, gasification, pyrolysis, torrefaction, hydrothermal liquefaction^{68,69}(Table 2).

Table 2: Second generation technologies

Process	Temperature & Pressure	Products
Thermochemical routes	150-3740C	Hydrogen, carbon monoxide, carbon dioxide, methane
Gasification	>700°C	Diesel, biomethanol, gasoline
Pyrolysis	430°C under pressure	Bio oil
Torrefaction	200-320°C	Bio oil
Hydrothermal Liquefaction	400°C and higher than atmospheric pressure	Bio oil, bio crude

1.4 CURRENT STRATEGIC PRODUCTION OF BIOFUEL

Biofuels obtain energy by the process of biological carbon fixation. United States leading the top most producer is

biofuels (1,557 petajoules/year) where, they have gradually increased from 187 thousand barrels per day in 2000 to 1.8 million barrels per day in 2019. A statistic representation of leading countries was published by Ian Tiseo, September 2, 2020 is represented in the Table 3.⁷⁰

Table 3: Biofuel production in developed countries

Countries	Biofuel production (Petajoules)
United States	1557.1
Brazil	992.2
Indonesia	275.5
Germany	143.4
France	113
China	111.3
Argentina	102.8
Thailand	95.6
Netherlands	79.2
Spain	66.7

The largest consumers of biofuels in US are National Army (Vehicles are fueled using 10% ethanol). US mainly uses corn (5.55 billion bushels in 2018) and soybeans for biodiesel production and according to Energy Information Administration (EIA) US produced 16.061 billion gallons of ethanol. Brazil which ranks second uses sugarcane and soybeans and the production 30.755 billion litres of ethanol in 2018 which was 9% more than 2017. Germany produced 75.8 thousand barrels/day in 2018

(2.9% of global biofuel production in 2018). ADM Olmuhle Hamburg (American group) is one of the major producer of biodiesel in Germany. Argentina leads the fourth global production (70.6 thousand barrels / day in 2018) which accounts for 2017% of world's production. There are 19 bioethanol plants of 1.4 billion litres/year production and sugarcane is the raw material in all these plants. Gela bio refinery in Europe which was launched in August 2019 utilizes second generation raw

materials and produced 7,50,000 tonnes annually of biofuel (70% of SO_2 , CO_2 , dust are reduced).

1.5 ALCOHOLIC FUELS

Aliphatic alcohols ($\text{C}_n\text{H}_{2n+1}\text{OH}$) such as methanol, ethanol, propanol and butanol are used as fuel for internal combustion

engines. The advantage of alcoholic fuels recorded is that they all have High Octane Rating which tends to increase their fuel efficiency. These alcoholic fuels are derived from fossil fuel and biomass and their Research Octane Number (RON) and Motor Octane Number (MON) and their energy density are tabulated in Table 4⁷¹.

Table: 4 Alcoholic fuel characteristics			
Fuel	Energy density (megajoules / Liter)	Research Octane Number (RON)	Motor Octane Number (MON)
Butanol	-30	96	78
Ethanol	-20	109	90
Propanol	-24	108	118
Methanol	-16	109	89

Alcoholic fuels have the potential to reduce NO_x (25-32%), CO (12-24%) and MC (20-22%) due to lower carbon to hydrogen ratio and improved engine efficiency. Propane and butanol are less toxic and less volatile than methanol. The cellulosic fermentation with *Clostridium acetobutylicum* processes produces propanol and butanol with extremely unpleasant smell. Swiss company (Butaico GmbH) adopts

modified yeast in the production of butanol and on combustion they give out CO_2 , water and heat. Ethanol has been used as rocket fuel and even in lightweight rocket powered racing aircraft and the energy content of ethanol is compared with other fuels is tabulated in Table 5⁷² and schematic representation of biofuel production in Fig.1⁷³

Table 5: Comparison efficiency of ethanol		
Fuel	Energy content (MJ/L)	Research Octane Number (RON)
Methanol	17.9	109
Ethanol	21.2	109
E85(85% ethanol + Gasoline 15%)	25.2	105
LPG	25.3	
Aviation gasoline	33.5	100
Gasohol	33.7	93
Regular gasoline	34.8	91
Diesel	38.5	25

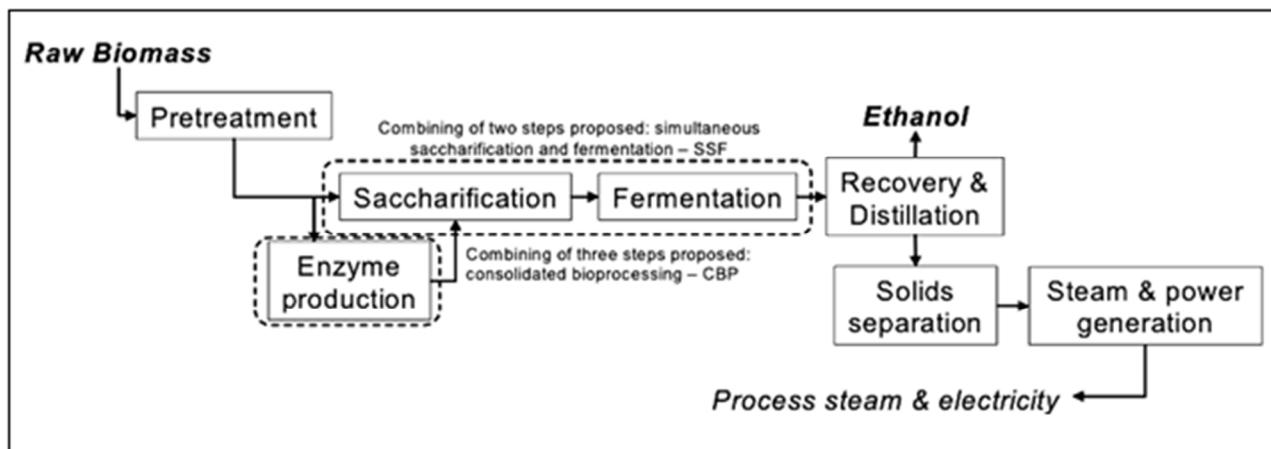


Fig 1: Schematic representation of ethanol production

1.6 ADVANCED BIOFUEL PRODUCTION THROUGH METABOLIC ENGINEERING

The demand in biofuel usage has raised the diversity of microorganisms in strain optimization in production of biofuel at industrial sectors in large scale. The engineered microorganisms (*E.coli*, *S. cerevisiae* and *Yarrowia lipolytica*) are more effective in the production of advanced biofuel from lignocellulosic biomass⁷⁴. In recent years isobutanol and n butanol has become more popular because of its low water

solubility, anti knock properties and energy densities⁷⁵. *Clostridium* strains are proven to produce butanol through acetone-butanol-ethanol (ABE) fermentation^{76,77}. Engineered *E.coli* has greater commercial implementation in which isobutanol pathway is altered to anaerobic conditions and secondly the cofactor preference for NADH over NADPH in ilvc and adhA^{78,79}. *E.coli* in another engineered method proved to increase the yield of isobutanol by excretion of ammonia and deleting both gdhA and glnA gene (used in nitrogen assimilation) in Erlich-like pyruvate pathway. Six fold

improvement of n butanol production (120 mg/L) was observed in heterologous expression of cytosolic pdh gene (inhibits ethanol and glycerol formation) using *S.cerevisiae*⁸⁰ and 242.8 mg/L of n butanol titers was reported in keto acid pathway by double deletion of *ilv2* and *adh1* by directing cytosolic pyruvate to mitochondrial threonine metabolism⁸². Two distinct pathways such as isopentenyl diphosphate (IPP) and dimethyl allyl diphosphate (DMAPP). And proved to be the precursors of isoprenoid fuels serving more effectively in jet engines⁸³. *E.coli* strains proved to be efficient in the isoprenoid derived branched C₅ alcohol production by overexpression of *mudF* gene and alleviating the rate limiting step of diphosphate hydrolysis⁸⁴. The purity of the product was improved when the pathway overexpressed the absence of *idi* gene in IPP to DMAPP⁸⁵. Another proven evidence is that in optimization of MVA pathway, when multiple -omics data analysis, coupled with ribosome binding site (optimization for *onudB*) increased the production to 2.2g/L with 1% glucose substratum^{86,87}. Conventional biodiesels were produced from plant oils and animal fats through fatty acid ethyl esters (FAEE) and fatty acid methyl esters (FAME) via trans estification of lipids. Fatty acid biosynthesis are more advantageous for its purities, improving performance etc. *E.coli*, *S.cerevisiae* and *Yarrowia lipolytica* (Oleaginous yeast) proved to be very efficient in fatty acid production of conventional biodiesel and petroleum based diesel⁸⁸. High yield of production was achieved by altering DNA binding affinity (response to acyl CoA) or by implementation of malonyl CoA (FapR) or supply of acetyl CoA and consumption of malonyl-CoA/ACP⁸⁹ or by linking upregulated FadfR or bypass ATP-dependent activation of acetyl CoA (C₂)⁹⁰ or optimized lipoylation pathway⁹¹.

5. REFERENCES

- Elshahed MS. Microbiological aspects of biofuel production: current status and future directions. *Advanced research*. 2010 Apr 1;1(2):103-11.
- Bhatt SM, Shilpa. Lignocellulosic feedstock conversion, inhibitor detoxification and cellulosic hydrolysis – a review. *Biofuels*. 2014 Nov 2;5(6):633-49. doi: 10.1080/17597269.2014.1003702.
- Wilderer PA. Global crises challenge environmental science and biotechnology. *Rev Environ Sci Bio Technol*. 2009 Dec 1;8(4):291-4. doi: 10.1007/s11157-009-9173-z.
- Chu S, Majumdar A. Opportunities and challenges for a sustainable energy future. *Nature*. 2012 Aug;488(7411):294-303. doi: 10.1038/nature11475, PMID 22895334.
- Hein KRG. Future energy supply in Europe-challenge and chances. *Fuel*. 2005 Jul 1;84(10):1189-94. doi: 10.1016/j.fuel.2004.09.022.
- Barnard D, Casanueva A, Tuffin M, Cowan D. Extremophiles in biofuel synthesis. *Environ Technol*. 2010 Jul 1;31(8-9):871-88. doi: 10.1080/09593331003710236, PMID 20662378.
- Dufey A. Biofuels production, trade and sustainable development: emerging issues. *lied*; 2006.
- Antoni D, Zverlov VV, Schwarz WH. Biofuels from microbes. *ApplMicrobiolBiotechnol*. 2007 Nov 1;77(1):23-35. doi: 10.1007/s00253-007-1163-x, PMID 17891391.
- Luque R, Herrero-Davila L, Campelo JM, Clark JH, Hidalgo JM, Luna D, Marinas JM, Romero AA. Biofuels:

2. CONCLUSION

Fossil fuels are limited and there may be a demand for coal, oil and natural gas and therefore the biofuels can work as an alternative form of fuel and reduce the pollution rate in the future. Biofuels are produced locally and therefore many countries have reduced their dependence on fossil fuels. Around more than 84% of the world's petroleum is utilized in the US and demand for transportation fuel is dramatically increasing day by day. The current scenario in the improvement in pretreatment, efficacy of enzymatic digestion, fermentation processes using efficient new strains will definitely increase the production of biofuel and strategic performance of eco friendly environment. Collective efforts in strain development, optimization process, novel biological pathways production rate and yield etc. will now be capable of developing economically feasible high yielding and efficient biofuels in industrial sectors. Apparent direction of recent advancement in metabolic engineering titers of these biofuel production requires significant improvement before commercial scale production.

3. AUTHORS CONTRIBUTION STATEMENT

Dr.R.Devika- Writing and compilation, M Subathra- Typing of manuscript, Dr. Madusudhan and others- Collection of articles.

4. CONFLICT OF INTEREST

Conflict of interest declared none.

- a technological perspective. *Energy Environ Sci*. 2008;1(5):542-64. doi: 10.1039/b807094f.
- Gerdy C, Glansdorff N. *Physiology and biochemistry of extremophiles*. ASM press; 2007.
- Sánchez OJ, Cardona CA. Trends in biotechnological production of fuel ethanol from different feedstocks. *Bioresour Technol*. 2008 Sep 1;99(13):5270-95. doi: 10.1016/j.biortech.2007.11.013, PMID 18158236.
- Braun R, Drosig B, Bochmann G, Weiß S, Kirchmair R. Recent developments in bio-energy recovery through fermentation. *Inmicrobes at Work* 2010 (pp. 35-58). Berlin, Heidelberg: Springer.
- Schmid J. Integration of biomass into future energy systems. In: *Proceedings of the 16th European biomass conference & exhibition*. Valencia: Spain; 2008 Jun 2-6.
- Gupta A, Verma JP. Sustainable bio-ethanol production from agro-residues: a review. *Renew Sustain Energy Rev*. 2015 Jan 1;41:550-67. doi: 10.1016/j.rser.2014.08.032.
- Cardona CA, Quintero JA, Paz IC. Production of bioethanol from sugarcane bagasse: status and perspectives. *Bioresour Technol*. 2010 Jul 1;101(13):4754-66. doi: 10.1016/j.biortech.2009.10.097, PMID 19945863.
- Singh A, Kuila A, Adak S, Bishai M, Banerjee R. Utilization of vegetable wastes for bioenergy generation. *Agric Res*. 2012 Sep 1;1(3):213-22. doi: 10.1007/s40003-012-0030-x.
- Lau MW, Dale BE. Cellulosic ethanol production from AFEX-treated corn stover using *Saccharomyces*

cerevisiae 424A (LNH-ST). *Proc Natl Acad Sci U S A*. 2009 Feb 3;106(5):1368-73. doi: 10.1073/pnas.0812364106, PMID 19164763.

18. Raghu S, Anderson RC, Daehler CC, Davis AS, Wiedenmann RN, Simberloff D, Mack RN. Adding biofuels to the invasive species fire?. *SCIENCE-New York THEN Washington*; 2006 (Sep 22): 313(5794): 1742.

19. Ingram LO, Conway T, Clark DP, Sewell GW, Preston JF. Genetic engineering of ethanol production in *Escherichia coli*. *Appl Environ Microbiol*. 1987 Oct 1;53(10):2420-5. doi: 10.1128/AEM.53.10.2420-2425.1987, PMID 3322191.

20. Gieg LM, Duncan KE, Suflita JM. Bioenergy production via microbial conversion of residual oil to natural gas. *Appl Environ Microbiol*. 2008 May 15;74(10):3022-9. doi: 10.1128/AEM.00119-08, PMID 18378655.

21. CerqueiraLeiteRCd, Verde Leal MRL, Barbosa Cortez LA, Griffin WM, Gaya Scandifio MI. Can Brazil replace 5% of the 2025 gasoline world demand with ethanol? *Energy*. 2009 May 1;34(5):655-61. doi: 10.1016/j.energy.2008.11.001.

22. Walter A, Cortez L. An historical overview of the Brazilian bioethanol program. *Renew Energy Dev*. 1999.

23. Rosillo-Calle F, Cortez LAB. Towards ProAlcool II—a review of the Brazilian bioethanol programme. *Biomass Bioenergy*. 1998 Mar 23;14(2):115-24. doi: 10.1016/S0961-9534(97)10020-4.

24. Bailey BK. Performance of ethanol as a transportation fuel. In *Handbook Bioethanol*. 2018 May 2:37-60.

25. Oliveira GdLT, McKay B, Plank C. How biofuel policies backfire: misguided goals, inefficient mechanisms, and political-ecological blind spots. *Energy Policy*. 2017 Sep 1;108:765-75. doi: 10.1016/j.enpol.2017.03.036.

26. LuckertMK. Market and government failures in the competition for land for biofuel production in Canada. *Biofuels*. 2014 May 4;5(3):211-8. doi: 10.1080/17597269.2014.913899.

27. Pacini H, Assunção L, Van Dam J, Toneto R. The price for biofuels sustainability. *Energy Policy*. 2013 Aug 1;59:898-903. doi: 10.1016/j.enpol.2013.03.042.

28. Pereira LG, Chagas MF, Dias MOS, Cavalett O, Bonomi A. Life cycle assessment of butanol production in sugarcane biorefineries in Brazil. *J Cleaner Prod*. 2015 Jun 1;96:557-68. doi: 10.1016/j.jclepro.2014.01.059.

29. Radhika K, Ravinder R, Ravindra P. Bioconversion of pentose sugars into ethanol: a review and future directions. *Biotechnol Mol Biol Rev*. 2011 Jan 31;6(1):8-20.

30. Chouinard-Dussault P, Bradt L, Ponce-Ortega JM, El-Halwagi MM. Incorporation of process integration into life cycle analysis for the production of biofuels. *Clean Technol Environ Policy*. 2011 Oct 1;13(5):673-85. doi: 10.1007/s10098-010-0339-8.

31. Biobutanol DP. an attractive biofuel. *Biotechnol J Healthc Nutr Technol*. 2007 Dec;2(12):1525-34.

32. Pedersen TH, Grigoras IF, Hoffmann J, Toor SS, Daraban IM, Jensen CU, Iversen SB, Madsen RB, Glasius M, Arturi KR, Nielsen RP, Søgaard EG, Rosendahl LA. Continuous hydrothermal co-liquefaction of aspen wood and glycerol with water phase recirculation. *Appl Energy*. 2016 Jan 15;162:1034-41. doi: 10.1016/j.apenergy.2015.10.165.

33. Yang J, He Q, Niu H, Corscadden K, Astatkie T. Hydrothermal liquefaction of biomass model components for product yield prediction and reaction pathways exploration. *Appl Energy*. 2018 Oct 15;228:1618-28. doi: 10.1016/j.apenergy.2018.06.142.

34. Dinita BJ, Malla SJ, Sreerama L. Lignocellulosic ethanol production: current practices and recent developments. *Biotechnol Mol Biol Rev*. 2011 Nov 30;6(8):172-82.

35. Saini JK, Saini R, Tewari L. Lignocellulosic agriculture wastes as biomass feedstocks for second-generation bioethanol production: concepts and recent developments. *3 Biotech*. 2015 Aug 1;5(4):337-53. doi: 10.1007/s13205-014-0246-5, PMID 28324547.

36. Wyman CE, Dale BE, Elander RT, Holtzapple M, Ladisch MR, Lee YY. Comparative sugar recovery data from laboratory scale application of leading pretreatment technologies to corn stover. *Bioresour Technol*. 2005 Dec 1;96(18):2026-32. doi: 10.1016/j.biortech.2005.01.018, PMID 16112491.

37. Deschamps FC, Ramos LP, Fontana JD. Pretreatment of sugar cane bagasse for enhanced ruminal digestion. *Appl Biochem Biotechnol*. 1996 Mar 1;57-58(1):171-82. doi: 10.1007/BF02941697, PMID 8669896.

38. Söderström J, Galbe M, Zacchi G. Separate versus simultaneous saccharification and fermentation of two-step steam pretreated softwood for ethanol production. *J Wood Chem Technol*. 2005 Jul 1;25(3):187-202. doi: 10.1080/02773810500191807.

39. Uellendahl H, Wang G, Møller HB, Jørgensen U, Skjadas IV, Gavala HN, Ahring BK. Energy balance and cost-benefit analysis of biogas production from perennial energy crops pretreated by wet oxidation. *Water Sci Technol*. 2008 Nov 1;58(9):1841-7. doi: 10.2166/wst.2008.504, PMID 19029727.

40. Vadas PA, Barnett KH, Undersander DJ. Economics and energy of ethanol production from alfalfa, corn, and switchgrass in the Upper Midwest, USA. *Bioenerg Res*. 2008 Mar 1;1(1):44-55. doi: 10.1007/s12155-008-9002-1.

41. Schöftner R, Valentin K, Schmidinger B, Trogisch S, Haberbauer M, Katzlunger K, Schnitzhofer W, Wernan N. Best Biogas Practise. Reports from energy and environmental research. 2007;45:2006.

42. Rosenberger A, Kaul H-P, Senn T, Aufhammer W. Improving the energy balance of bioethanol production from winter cereals: the effect of crop production intensity. *Appl Energy*. 2001 Jan 1;68(1):51-67. doi: 10.1016/S0306-2619(00)00036-2.

43. Bazmi AA, Bhutto AW, Ghauri M. Ethanol fuel as feasible and desired options in Pakistan. *ESdev-ClIT Abbottabad*, Pakistan. 2007.

44. Warren RK, Macdonald DG, Hill GA. The design and costing of a continuous ethanol process using wheat and cell recycle fermentation. *Bioresour Technol*. 1994 Jan 1;47(2):121-9. doi: 10.1016/0960-8524(94)90109-0.

45. Jones AM, Thomas KC, Inglede W. Ethanolic fermentation of blackstrap molasses and sugarcane juice using very high gravity technology. *J Agric Food Chem*. 1994 May;42(5):1242-6. doi: 10.1021/jf00041a037.

46. Vander Stichele M, Bizzarri K, Plank L. Corporate power over EU trade policy: good for business, bad for the world. *Brussels: Seattle to Brussels Network*; 2006.

47. Mariano AP, Dias MO, Junqueira TL, Cunha MP, Bonomi A, Filho RM. Butanol production in a first-generation Brazilian sugarcane biorefinery: technical aspects and economics of greenfield projects. *Bioresour Technol.* 2013 May 1;135:316-23. doi: 10.1016/j.biortech.2012.09.109, PMID 23127845.

48. Kumar M, Gayen K. Developments in biobutanol production: new insights. *Appl Energy.* 2011 Jun 1;88(6):1999-2012. doi: 10.1016/j.apenergy.2010.12.055.

49. Lopes ML, Paulillo SC, Godoy A, Cherubin RA, Lorenzi MS, Giometti FH, Bernardino CD, AmorimNeto HB, Amorim HV. Ethanol production in Brazil: a bridge between science and industry. *Braz J Microbiol.* 2016 Dec 1;47 Suppl 1:64-76. doi: 10.1016/j.bjm.2016.10.003, PMID 27818090.

50. Birkay CW. The inheritance of genes in mitochondria and chloroplasts: laws, mechanisms, and models. *Annu Rev Genet.* 2001 Dec;35(1):125-48. doi: 10.1146/annurev.genet.35.102401.090231, PMID 11700280.

51. Benítez T, Martínez P, Codón AC. Genetic constitution of industrial yeast. *Microbiología.* 1996 Sep;12(3):371-84. PMID 8897417.

52. Vezinhet F, Blondin B, Hallet JN. Chromosomal DNA patterns and mitochondrial DNA polymorphism as tools for identification of enological strains of *Saccharomyces cerevisiae*. *Appl Microbiol Biotechnol.* 1990 Feb 1;32(5):568-71. doi: 10.1007/BF00173729.

53. Panek AC, Vânia JJ, Paschoalin MF, Panek D. Regulation of trehalose metabolism in *Saccharomyces cerevisiae* mutants during temperature shifts. *Biochimie.* 1990 Jan 1;72(1):77-9. doi: 10.1016/0300-9084(90)90176-h, PMID 2160289.

54. Heinze T, Dicke R, Koschella A, Kull AH, Klohr EA, Koch WV. Effective preparation of cellulose derivatives in a new simple cellulose solvent. *Macromol Chem Phys.* 2000 Mar 1;201(6):627-31. doi: 10.1002/(SICI)1521-3935(20000301)201:6<627::AID-MACP627>3.0.CO;2-Y.

55. Dombek KM, Ingram LO. Ethanol production during batch fermentation with *Saccharomyces cerevisiae*: changes in glycolytic enzymes and internal pH. *Appl Environ Microbiol.* 1987 Jun 1;53(6):1286-91. doi: 10.1128/AEM.53.6.1286-1291.1987, PMID 3300550.

56. Gumel AM, Idris A, Wada-Kura A, Ibrahim MM, Mustapha IU. Turning Waste to Wealth: A mini review on Bioethanol Production from Renewable Biomass.

57. Brandt A, Gräsvik J, Hallett JP, Welton T. Deconstruction of lignocellulosic biomass with ionic liquids. *Green Chem.* 2013;15(3):550-83. doi: 10.1039/c2gc36364j.

58. Binod P, Sindhu R, Singhania RR, Vikram S, Devi L, Nagalakshmi S, Kurien N, Sukumaran RK, Pandey A. Bioethanol production from rice straw: an overview. *Bioresour Technol.* 2010 Jul 1;101(13):4767-74. doi: 10.1016/j.biortech.2009.10.079, PMID 19944601.

59. Sun Y, Cheng J. Hydrolysis of lignocellulosic materials for ethanol production: a review. *Bioresour Technol.* 2002 May 1;83(1):1-. doi: 10.1016/s0960-8524(01)00212-7, PMID 12058826.

60. Sarkar N, Ghosh SK, Bannerjee S, Aikat K. Bioethanol production from agricultural wastes: an overview. *Renew Energy.* 2012 Jan 1;37(1):19-27. doi: 10.1016/j.renene.2011.06.045.

61. Tengborg C, Galbe M, Zacchi G. Influence of enzyme loading and physical parameters on the enzymatic hydrolysis of steam-pretreated softwood. *Biotechnol Prog.* 2001;17(1):110-7. doi: 10.1021/bp000145+, PMID 11170488.

62. Zhu S, Wu Y, Chen Q, Yu Z, Wang C, Jin S, Ding Y, Wu G. Dissolution of cellulose with ionic liquids and its application: a mini-review. *Green Chem.* 2006;8(4):325-7. doi: 10.1039/b601395c.

63. Sarker TC, Azam SMGG, Bonanomi G. Recent advances in sugarcane industry solid by-products valorization. *Waste Biomass Valorization.* 2017 Mar 1;8(2):241-66. doi: 10.1007/s12649-016-9665-3.

64. Pandey A, Soccol CR, Nigam P, Soccol VT. Biotechnological potential of agro-industrial residues. I: sugarcane bagasse. *Bioresour Technol.* 2000 Aug 1;74(1):69-80. doi: 10.1016/S0960-8524(99)00142-X.

65. Gupta A, Verma JP. Sustainable bio-ethanol production from agro-residues: a review. *Renew Sustain Energy Rev.* 2015 Jan 1;41:550-67. doi: 10.1016/j.rser.2014.08.032.

66. Khatiwada D, Leduc S, Silveira S, McCallum I. Optimizing ethanol and bioelectricity production in sugarcane biorefineries in Brazil. *Renew Energy.* 2016 Jan 1;85:371-86. doi: 10.1016/j.renene.2015.06.009.

67. Inderwildi OR, King DA. Quo vadis biofuels?. *Energy Environ Sci.* 2009;2(4). doi: 10.1039/b822951c.

68. Ramirez J, Brown R, Rainey T. A Review of hydrothermal liquefaction bio-crude properties and prospects for upgrading to transportation fuels. *Energies.* Jul 2015;8(7):6765-94. doi: 10.3390/en8076765.

69. Zhou H, Long Y, Meng A, Chen S, Li Q, Zhang Y. A novel method for kinetics analysis of pyrolysis of hemicellulose, cellulose, and lignin in TGA and macro-TGA. *RSC Adv.* 2015;5(34):26509-16. doi: 10.1039/C5RA02715B.

70. Zhou H, Long YQ, Meng AH, Li Q, Zhang Y. The pyrolysis simulation of five biomass species by hemicellulose, cellulose and lignin based on thermogravimetric curves. *Thermochim Acta.* Aug 2013;566:36-43. doi: 10.1016/j.tca.2013.04.040.

71. Liebig J. On the constitution of ether and its compounds. *Annu Pharm.* 2004;9(22):1-39.

72. Turner C, Spanel P, Omith. A longitudinal study of ethanol and acetaldehyde in the exhaled breath of healthy volunteers using selected ion flow tube mass spectroscopy. *Rapid Commun Mass Spectrom.* 2002;20(1):61-8.

73. Schenk PM, Thomas-Hall SR, Stephens E, Marx UC, Mussgnug JH, Posten C, Kruse O, Hankamer B. Second generation biofuels: high-efficiency microalgae for biodiesel production. *Bioenerg Res.* 2008 Mar 1;1(1):20-43. doi: 10.1007/s12155-008-9008-8.

74. Rothamer DA, Donohue TJ. Chemistry and combustion of fit-for-purpose biofuels. *Curr Opin Chem Biol.* 2013;17(3):522-8. doi: 10.1016/j.cbpa.2013.03.039, PMID 23664492.

75. Jones DT, Woods DR. Acetone-butanol fermentation revisited. *Microbiol Rev.* 1986;50(4):484-524. doi: 10.1128/MMBR.50.4.484-524.1986, PMID 3540574.

76. Rabinovitch-Deere CA, Oliver JWK, Rodriguez GM, Atsumi S. Synthetic biology and metabolic engineering approaches to produce biofuels. *Chem Rev.*

2013;113(7):4611-32. doi: 10.1021/cr300361t, PMID 23488968.

77. Atsumi S, Cann AF, Connor MR, Shen CR, Smith KM, Brynildsen MP, Chou KJ, Hanai T, Liao JC. Metabolic engineering of *Escherichia coli* for 1-butanol production. *Metab Eng.* 2008;10(6):305-11. doi: 10.1016/j.ymben.2007.08.003, PMID 17942358.

78. Harvey BG, Meylmans HA. The role of butanol in the development of sustainable fuel technologies. *J Chem Technol Biotechnol.* 2011;86(1):2-9. doi: 10.1002/jctb.2540.

79. Atsumi S, Wu TY, Eckl EM, Hawkins SD, Buelter T, Liao JC. Engineering the isobutanol biosynthetic pathway in *Escherichia coli* by comparison of three aldehyde reductase/alcohol dehydrogenase genes. *Appl Microbiol Biotechnol.* 2010;85(3):651-7. doi: 10.1007/s00253-009-2085-6.

80. Bastian S, Liu X, Meyerowitz JT, Snow CD, Chen MMY, Arnold FH. Engineered ketol-acid reductoisomerase and alcohol dehydrogenase enable anaerobic 2-methylpropan- 1-ol production at theoretical yield in *Escherichia coli*. *Metab Eng.* 2011;13(3):345-52. doi: 10.1016/j.ymben.2011.02.004.

81. Shen CR, Lan El, Dekishima Y, Baez A, Cho KM, Liao JC. Driving forces enable high-titer anaerobic 1-butanol synthesis in *Escherichia coli*. *Appl Environ Microbiol.* 2011;77(9):2905-15. doi: 10.1128/AEM.03034-10.

82. Bond-Watts BB, Bellerose RJ, Chang MCY. Enzyme mechanism as a kinetic control element for designing synthetic biofuel pathways. *Nat Chem Biol.* 2011;7(4):222-7. doi: 10.1038/nchembio.537, PMID 21358636.

83. Baez A, Cho KM, Liao JC. High-flux isobutanol production using engineered *Escherichia coli*: a bioreactor study with in situ product removal. *Appl Microbiol Biotechnol.* 2011;90(5):1681-90. doi: 10.1007/s00253-011-3173-y, PMID 21547458.

84. Huo YX, Cho KM, Rivera JG, Monte E, Shen CR, Yan Y, Liao JC. Conversion of proteins into biofuels by engineering nitrogen flux. *Nat Biotechnol.* 2011;29(4):346-51. doi: 10.1038/nbt.1789, PMID 21378968.

85. Xu P, Li L, Zhang F, Stephanopoulos G, Koffas M. Improving fatty acids production by engineering dynamic pathway regulation and metabolic control. *Proceedings of the National Academy of Sciences.* 2014;111(31):11299-304. doi: 10.1073/pnas.1406401111.

86. Zhang F, Carothers JM, Keasling JD. Design of a dynamic sensor-regulator system for production of chemicals and fuels derived from fatty acids. *Nat Biotechnol.* 2012;30(4):354-9. doi: 10.1038/nbt.2149.

87. Liu H, Yu C, Feng DX, Cheng T, Meng X, Liu W, Zou H, Xian M. Production of extracellular fatty acid using engineered *Escherichia coli*. *Microb Cell Factories.* 2012;11(1). doi: 10.1186/1475-2859-11-41.

88. Dellomonaco C, Clomburg JM, Miller EN, Gonzalez R. Engineered reversal of the beta-oxidation cycle for the synthesis of fuels and chemicals. *Nature.* 2011;476(7360):355-9. doi: 10.1038/nature10333.

89. Goh EB, Baidoo EE, Keasling JD, Beller HR. Engineering of bacterial methyl ketone synthesis for biofuels. *Appl Environ Microbiol.* 2012;78(1):70-80. doi: 10.1128/AEM.06785-11, PMID 22038610.

90. Guo D, Zhu J, Deng Z, Liu T. Metabolic engineering of *Escherichia coli* for production of fatty acid short-chain esters through combination of the fatty acid and 2-keto acid pathways. *Metab Eng.* 2014;22:69-75. doi: 10.1016/j.ymben.2014.01.003.

91. Cao YX, Xiao WH, Liu D, Zhang JL, Ding MZ, Yuan YJ. Biosynthesis of odd-chain fatty alcohols in *Escherichia coli*. *Metab Eng.* 2015;29:113-23. doi: 10.1016/j.ymben.2015.03.005, PMID 25773521.