



Phytochemical Assessment of Certain Seaweeds of Enteromorpha Species in Coastal Regions of Ramanathapuram, Tamil Nadu, India

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Abstract: Freshly dried seaweeds are being extensively consumed, particularly by peoples living in coastal areas. *Enteromorpha* algae are nutritious and low-calorie food rich in essential Amino acids, Fatty acids, Vitamins, Dietary fiber and Resistant protein of humans. *E. prolifera* has many nutritional compounds and polysaccharides that have attracted extensive interest due to their numerous biological activities. The current studies for uses some Phytochemical components such as Alkaloids, Flavonoids, Phenols, Proteins, free Amino acids, Saponins, Sterols, Terpenoids, Coumarin, Glycosides, Quinones and Tannins are evaluated on the chosen *Enteromorpha intestinalis* species, including *E. flexuosa*, *E. intestinalis* and *E. prolifera*. In addition, Protein, Carbohydrate, Fat and photosynthetic pigments like chlorophyll and carotenoids were also quantitatively determined.

Keywords: *Enteromorpha* species, Phytochemical Assessment, Selected seaweeds.

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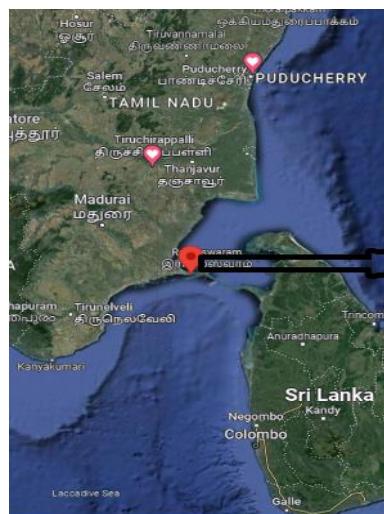
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I. INTRODUCTION

Algae is known as seaweeds, which are classified permitting to their natural pigments; green, brown, and red seaweeds. Seaweeds are recognized as a source of bioactive compounds.¹⁶ The numbers of metabolites considered by a broad spectrum of biological activities and traditionally used as nutritive specific taste foods in Asia. The freshly dried seaweeds are being extensively consumed, particularly by people living in coastal areas¹³. The invention of valuable chemical compounds; Alginates, Agar, Carrageenan and polysaccharides¹¹. These seaweeds are used in food, pharmaceutical and other industries to enhance the nutritional value of food, stabilize emulsions and make products more palatable¹⁹. They are also used to thicken and stabilize food products, improve the texture of ice cream, and even as a health supplement due to their high concentration of vitamins and minerals²⁰. These compounds are known as emulsifiers and they work by binding oil and water molecules together, creating a homogenous mixture. This allows for the uniform dispersion of fat-soluble vitamins and minerals, improving the product's overall nutritional value²¹. They also help to prevent spoilage and extend the shelf life of food products²². *Enteromorpha* is nutritious and low-calorie food rich in essential Amino acids, Fatty acids, Vitamins, Dietary fiber and Resistant protein of humans⁸. *Enteromorpha* also contains chlorophyll b and other minerals like Calcium, Magnesium, Iron, etc., Chemical analysis exhibited that *Enteromorpha* species have protein, ash content and total Fatty acids are respectively. They stated that polysaccharides like carbohydrate polymers are used in food, cosmetic and pharmaceutical industries, microbiology, and biotechnology. This type of polysaccharide is a major property of *E. prolifera*. *E. prolifera* has many nutritional compounds and polysaccharides that have attracted extensive interest due to their numerous biological activities. Polysaccharide compounds use plays a nutritional role as dietary fiber and biological activities like gelling abilities. They stated that *Caulerpa serrulata* and *E. prolifera* were similar percentages of protein which equals that of cereals like oats. *E. intestinalis*- Sulfated polysaccharide reduced tumor mass; increased thymus and spleen mass; increased TNF- α , NO, and reactive oxygen species. *intestinalis* - Methanol extract Antiproliferative¹⁷.



Credit and Source: Google Maps



The Mandapam google satellite map. This place is situated in Ramanathapuram, Tamil Nadu, India, its geographical coordinates are 9° 17' 0" North, 79° 7' 0" East, and its original name are Mandapam.

The findings suggest that *Caulerpa serrulata* and *E. prolifera* contain proteins that are comparable in quality to those found in cereals like oats. Furthermore, they demonstrate that the sulfated polysaccharide in *E. intestinalis* has the potential to reduce tumor mass and increase thymus and spleen mass. In contrast, the methanol extract of this species has shown anti-proliferative activity. Moreover, the study highlights the potential therapeutic effects of these seaweeds, suggesting they could be beneficial in the treatment of cancer and other related diseases²³. However, more research is needed to determine the long-term effects of these seaweeds on cancer and other diseases. Additionally, it is unclear whether the benefits of these seaweeds outweigh the risks. *E. compressa* was observed in Mandapam coast regions Vasantha and Rajamanikam (2003)⁵⁻¹⁷ are reported the differentiation in the chemical constituents of the marine red algae in Tuticorin and Mandapam coast found out the nutritional properties of 20 species of seaweeds from the Gulf of Mannar². The present study carried out the analysis of phytochemical and biochemical pigments such as four marine seaweeds; *E. compressa* (Linnaeus) Nees, *E. flexuosa* (Wulfen) J. Agardh, *E. intestinalis* (Linnaeus) Nees and *E. prolifera* (Müller) and J. Agardh are belonging to the family Ulvaceae.

2. MATERIALS AND METHODS

2.1. Sample collection

Fresh plants of *E. compressa*, *E. flexuosa*, *E. intestinalis*, and *E. prolifera* were collected from the intertidal regions of Ramanathapuram District of Tamil Nadu, India and then they were brought to the laboratory. The algae were washed thoroughly with tap water to remove extraneous materials. The well-washed samples were dried in the oven at 37°C till constant weight. The dried samples were individually ground in an electric mixer.⁷ The powdered samples were then stored in the refrigerator until use. Plant Authentication is done by P. Sona, Research scholar & Dr. G. Subramanian, Head of the Department of Botany, Arignar Anna Govt. Arts College, Namakkal. The plants were stored with voucher numbers AAGACNTN021 (*Enteromorpha compressa*), AAGACNTN022 (*E. flexuosa*), AAGACNTN023 (*E.intestinalis*), AAGACNTN024 (*E.Prolifera*).

2.2. Distribution

It can be found in the Bering Sea near Alaska, the Aleutian Islands, Puget Sound, Japan, Korea, Mexico, the Philippines, and Russia. Besides this, it can be found in Israel and European countries such as the Azores, Belgium, Denmark, Ireland, Norway, Poland and the Baltic and Mediterranean Seas. It is also found on the Pacific Ocean's shores, including in New Zealand.

2.3. Description

The fronds have branches are completely tubular, expanding in width to mid-thallus, reaching 15 cm long or more. The cells are irregularly arranged, and the chloroplast is hood-shaped and placed to one side, generally with only one pyrenoid. The species may be 10–30 cm (3.9–11.8 in) long and 6–18 millimeters (0.24–0.71 in) wide. They have rounded tips as well. The algae may be reproductive at all times of the year and has a life cycle with an alternation of generations. The gametophyte and saprophytes are isomorphic, having identical morphology. In some references, the species (*Ulva intestinalis*) is treated as two subspecies: ssp. *intestinalis* (L.) Link and ssp. *Compressa*(L.)^{34,35}.

2.4. Medicinal properties of Enteromorpha

Enteromorpha is used against goiter and scrofula as an antipyretic to prepare a refreshing liquid and treat sunstroke, bronchitis, cough, and asthma. It is also applied as fish bait. In addition, the dried and crushed fronds can be used as a topping for many foods, in soups, and as a coating. *Enteromorpha* is rich in nutrients with medicinal and health-promoting effects. From a nutritional standpoint, the main properties of sea lettuce are its richness in polysaccharides, protein and Amino acids, Fatty acids, Minerals and Vitamins. Therefore, their nutritional value makes them valuable food supplements^{36,37}.

2.5. Phytochemical screening

The stored samples were subjected to a qualitative test to examine phytochemical content with the standard procedures⁴. The seaweed powder extracts were tested for Alkaloids, Flavonoids, Glycosides, Phenolic groups, Sugars, Saponins, Steroids, Tannins, Protein, Amino acids and Terpenoids.

2.6. Preparation of extracts³⁸

The seaweed specimens were washed thoroughly, placed on blotting paper, and spread out at 37 ±2°C in the shade condition for drying. Next, the shade-dried samples were grounded to a fine powder using a tissue blender. The powdered samples were then stored in the refrigerator for further use. Finally, 3g powdered samples were packed in the Soxhlet apparatus and extracted with ethanol for 12 hours.

- 1. Test for Alkaloids:** 1ml of 1% HCl was added to the 2ml extract in a test tube and treated with a few drops of Mayer's reagent. A creamy white precipitate indicated the presence of alkaloids.
- 2. Test for Flavonoids:** Five drops of 1% NH₃ solution were added to 2 ml of extract in a test tube. A yellow coloration was observed for the presence of flavonoids.
- 3. Test for Glycosides:** 2ml of 50% H₂SO₄ was added to

the 2ml of extract in a boiling tube. The mixture was heated in a boiling water bath for 5 min. Next, 10 ml of Fehling's solution was added and boiled. A brick-red precipitate indicated the presence of glycosides.

- 4. Test for Phenolic groups:** To 1ml extract, add 2ml distilled water followed by a few drops of 10% Ferric chloride. Either blue or black color formed showed the presence of phenolic groups.
- 5. Test for sugars:** 5-8 drops of Fehling's solution was added to 2ml extract. The mixture was heated in a boiling water bath for 5 min. A precipitate formed with either red or brick color showed the presence of sugar.
- 6. Test for Saponins:** About 2 ml of the extract was shaken vigorously with 5 ml distilled water to obtain stable, persistent foam. The formation of emulsion indicated the presence of saponins.
- 7. Test for Steroids:** About 0.5 ml of hot acetic anhydride was added with 2 ml of ethanol extract. The mixture was treated with Libermann reagent. The appearance of a ring of blue-green showed the presence of sterol and steroids.
- 8. Test for Tannins:** 1 ml of distilled water and 1-2 drops of ferric chloride solution were added in 2 ml extract and observed for brownish green or a blue-black coloration.
- 9. Test for Terpenoids:** 2ml of CHCl₃ was added in 2ml extract in a test tube. And then, 3 ml of concentrated H₂SO₄ was carefully added along the test tube wall to form a layer. An interface with a reddish-brown coloration has confirmed the presence of terpenoids.
- 10. Test for Coumarin:** For coumarin identification, 1 ml of extract and 1 ml of 10% NaOH were added. The formation of the yellow color indicates the presence of coumarins.
- 11. Test for Quinines:** For quinine identification, 1ml of extract, and 1ml of concentrated sulphuric acid were added. The formation of red color indicated the presence of quinones.
- 12. Test for Proteins:** To a 2ml of ethanolic extract, 1ml of 40% NaOH solution was added, and two drops of 1% CuSO₄ solution were added. The presence of a peptide linkage of the molecule was indicated by the violet color, which showed the presence of protein.
- 13. Test for Amino Acids:** To 2 ml of ethanolic extract, 2 ml of Ninhydrin reagent was added and laid in a water bath for about 20 minutes. The visual aspect of the purple color formed indicated the presence of amino acids.

2.7. Estimation of total Protein content

The protein was estimated by the Biurette method (Raymont et al., 1964)¹². To 2 mg of dried algal powdered sample, 2 ml of distilled water followed by 4 ml of burette reagent were added and incubated for 30 minutes at room temperature. Then the mixture was centrifuged for 15 minutes at 4000 rpm. Finally, the supernatant solution was pooled, and the optical density was taken at 540 nm in a spectrophotometer.

2.8. Estimation of total Lipid content

The lipid estimation was carried out using a solvent mixture of chloroform-methanol (2:1). 3 10 mg of dried algal powder sample was taken in a test tube, and 5 ml of chloroform-methanol (2:1) mixture was added. The test tubes with the mixture were closed with aluminum foil and then incubated at 37 ± 2°C for 24 hrs. After the incubation, the sample mixture was filtered using a whatman No.1 filter paper. The filtrate was collected and pooled in a 50 ml pre weighed

beaker, which was kept on a hot plate until the solvent evaporated. The residue with the beaker was weighed and calculated to know the total crude lipid of the sample.

2.9. Estimation of total Sugar content

The algal sample's total sugar content was estimated using the method described by¹⁴. A known quantity of the sample was taken in pestle and mortar, added 80% ethanol was, and then ground well and centrifuged at 4000 rpm. About 5 ml of the supernatant was taken in a test tube; 5 ml of anthrone reagent was added. The tube was kept in a boiling water bath for 20 min. After that, it was kept in the darkroom for another 20 minutes and then read in a spectrophotometer at 650 nm.

2.10. Estimation of total Chlorophyll content

The amount of chlorophyll present in the freshly collected seaweed was analyzed by the method of Arnon¹. About 500 mg of a fresh sample of seaweed was kept in a pestle and mortar with an adequate amount of 80% acetone, and then it was ground well. The homogenate liquid was centrifuged at 3000 rpm for 10 minutes, and the supernatant was stored. Next, the pellet was extracted by repeated washing with 80 % acetone until it turned colorless. Then, the extracts were pooled and subjected to determine the chlorophyll content. The extract absorbance was observed at 645 nm and 663 nm in a spectrophotometer. The chlorophyll content was calculated by using the following formula:

$$\text{Chlorophyll 'a' (mg/g.fr.wt.)} = \frac{12.7 \times A663 - 2.69 \times A645}{a \times 1000 \times W} \times V$$

$$\text{Chlorophyll 'b' (mg/g.fr.wt.)} = \frac{22.9 \times A645 - 4.68 \times A663}{a \times 1000 \times W} \times V$$

$$\text{Total Chlorophyll (mg/g.fr.wt.)} = \frac{20.2 \times A645 + 8.02 \times A663}{a \times 1000 \times W} \times V$$

Where A = Absorbance at respective wavelength = Volume of extract (ml), W = Fresh weight of the sample (g)

2.11. Estimation of total Carotenoid content

The same algal chlorophyll extract was used to measure carotenoids. The extract OD was taken at 480 nm⁶ Carotenoid (μg/g.fr.wt.) = A.480 + (0.114 X A. 663) – (0.638 X A. 645).

Where A = Absorbance at the respective wavelength

3. RESULTS AND DISCUSSION

3.1. Qualitative Test of Phytochemicals analysis

In this study, 12 important phytochemicals, namely Alkaloids,

Glycosides, Coumarins, Flavonoids, Phenols, Protein and amino acids, quinones, saponins, sterols, sugar, tannin and terpenoids were qualitatively tested in the selected four species of a genus *Enteromorpha* (Table -I). Among the 12 phytochemicals, coumarin, Glycosides, Quinone and Tannin were absent, and the remaining eight phytochemicals were present in the selected four species of *Enteromorpha* (Table-I). The brownest and red algal seaweeds have agar, alginic acid, laminarin, fucoidan, galactans, carrageenan, xylan, and mannans naturally, but in the green algal seaweeds, mostly the presence of alkaloids, flavonoids, phenol, protein and amino acid, sterols, sugar, and terpenoids. Some phytochemicals like Coumarin, Glycosides, Quinone and tannin were absent in all four algal samples (Table-I)

Table I: Qualitative Phytochemical Screening of Marine Enteromorpha Species				
Phytochemicals	<i>E. compressa</i>	<i>E. flexuosa</i>	<i>E. intestinalis</i>	<i>E. prolifera</i>
Alkaloids	+++	+++	++	+
Coumarin	-	-	-	-
Flavonoids	+++	+++	+	+++
Glycosides	-	-	-	-
Phenol	++	+++	+++	++
Protein and Amino acid	+++	++	++	+
Quinone	-	-	-	-
Saponin	+	+++	++	++
Sterols	+++	++	++	+
Sugar	+	+++	+	+++
Tannin	-	-	-	-
Terpenoids	+++	++	+	+++

Note: +++ (High), ++ (Moderate) - (Absent)

In the above table (Table 1), Alkaloids, Flavonoids, Phenol, Protein and Amino acids, Saponins, Sterols, Sugar and Terpenoids were present in all four species. But Coumarin Glycosides, Quinone, and Tannin were absent in all four species.

3.2. Estimation of Biochemicals

Among the four species, alkaloid was present in all the selected four species; *E. compressa* had the maximum Alkaloid of 2.84% than the remaining three species (Table -2) ²⁴. Flavonoid was also present in all four species of *Enteromorpha*, with a range of 1.33 to 1.63%. It is similar to early reports ²⁵⁻²⁷. The phenol compound was present maximally in *E. intestinalis* with 0.66% and a minimum of 0.22% in *E. flexuosa* (Table -2). Biochemical protein, sugar, and the lipid content of the selected four green algal species

of *Enteromorpha* were quantitatively estimated (Table -2). The Sugar, Protein and Lipids were maximally 7.94, 6.72, and 2.09 % in *E. prolifera*, respectively. Protein and sugar were minimally 6.78 and 5.77 %, respectively, in *E. compressa*, and lipid was 1.09% in *E. flexuosa*, less than the remaining three species (Table -2; Figures: 1- 4)²⁸. This present four species results showed similarity with the early reports⁹. The present study recorded the maximum protein content in the green alga *E. prolifera* and the minimum in the green alga *E. compressa*. Similarly, recorded the highest protein content in brown algae *Tubinaria ornata* from the Gulf of Mannar regions near Rameswaram². These reports support the present study that green algal seaweeds equal brown algal seaweed and red algal seaweeds. The present study showed the lipid content of *Enteromorpha* species was significantly equal to green algae *C. adhaerens* and *U. fasciata*.¹⁰

Table 2: Protein, Sugar, and lipid contents of marine algal species of enteromorpha

Biochemical (Result in%)	<i>E. compressa</i>	<i>E. flexuosa</i>	<i>E. intestinalis</i>	<i>E. prolifera</i>
Alkaloids	2.84 ±0.02	2.74 ± 0.01	2.64 ±0.05	2.06 ± 0.04
Flavonoids	1.42 ±0.05	1.33 ± 0.04	1.63 ±0.08	1.43 ± 0.05
Lipids	1.30 ±0.02	1.09 ± 0.11	1.42 ±0.31	2.09 ± 0.05
Phenols	0.33 ±0.02	0.22 ± 0.02	0.63 ±0.05	0.52 ± 0.07
Proteins	6.75 ±0.22	7.20 ± 0.04	7.52 ±0.15	7.94 ± 0.02
Sugar	5.74 ±0.01	6.52 ± 0.03	5.85 ±0.23	6.72 ± 0.14

All values are expressed in percentage; ± mean value represents standard deviation.

Alkaloids were found higher in *E. compressa*, Flavonoids higher in *E. intestinalis*, Lipids higher in *E. prolifera*, Phenols higher in *E. intestinalis* and Proteins higher in *E. prolifera*.

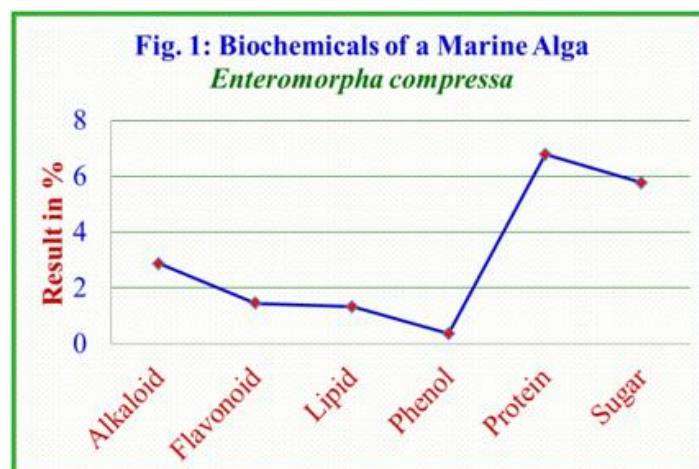


Fig 1: Biochemicals of a Marine Alga *Enteromorpha compressa*

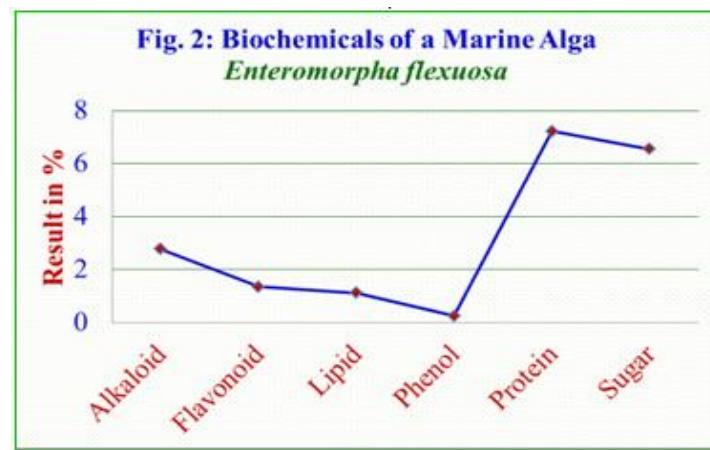


Fig 2: Biochemicals of a Marine Algae *Enteromorpha flexuosa*

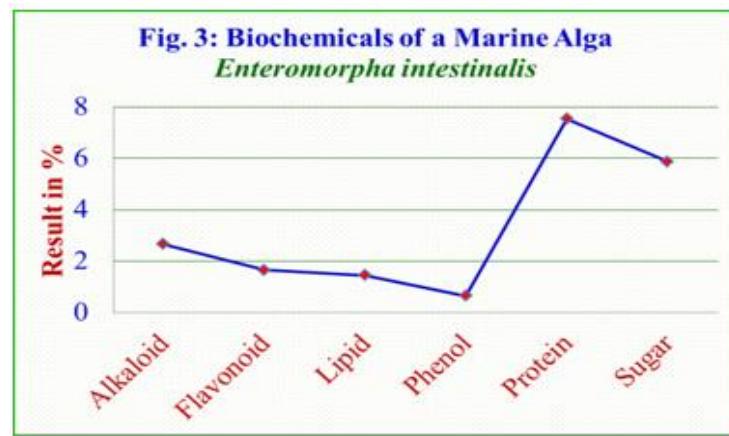


Fig 3: Biochemicals of a Marine Algae *Enteromorpha intestinalis*

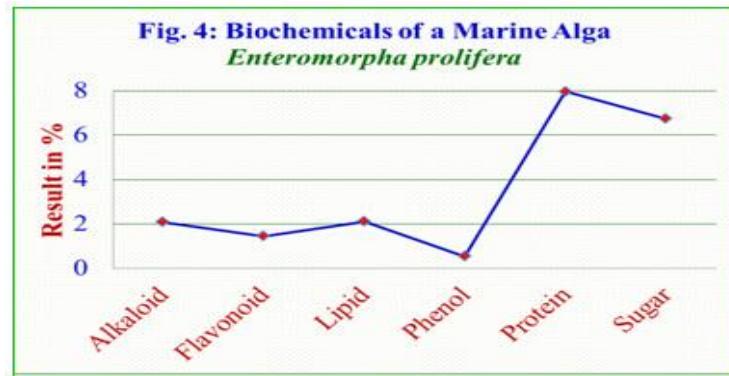


Fig 4: Biochemicals of a Marine Algae *Enteromorpha prolifera*

The sugar, protein, and lipid were maximally 7.94, 6.72, and 2.09 % in *E. prolifera*. Protein and sugar were minimally 6.78, and 5.77 %, respectively, in *E. compressa*, and lipid was 1.09% in *E. flexuosa*, which was less than the remaining three species (Table -2; Figures: 1- 4)

Table.3: pigments of four marine algal species of enteromorpha.				
AlgalPigments (mg/g offr.wt.)	<i>E. Compressa</i>	<i>E. flexuosa</i>	<i>E. Intestinalis</i>	<i>E. prolifera</i>
Chlorophyll a	0.344 ± 0.031	0.312 ± 0.021	0.391 ± 0.025	0.350 ± 0.052
Chlorophyll b	0.112 ± 0.011	0.124 ± 0.022	0.095 ± 0.023	0.088 ± 0.015
TotalChlorophyll	0.459 ± 0.042	0.439 ± 0.043	0.489 ± 0.048	0.441 ± 0.067
Carotenoids(µg/g.fr.wt)	0.372 ± 0.022	0.383 ± 0.025	0.319 ± 0.011	0.392 ± 0.255

All values are expressed in percentage; ± mean value represents the standard deviation.

The table revealed that algal pigments like chlorophyll 'a', chlorophyll 'b', total chlorophyll and Carotenoid were

present in all four species of Enteromorpha and they were estimated quantitatively with the highest chlorophyll a in *E.*

intestinalis with 0.388 ± 0.022 mg/gfr.wt., and the highest chlorophyll b in *E. flexuosa* with 0.124 mg/g offr. Wt, which is similar to some earlier reports^{18,29,30,31}. The species *E. intestinalis* had the highest total chlorophyll (0.492mg/g of fr.wt.), and *E. prolifera* had the maximum of Carotenoid ($0.395 \mu\text{g/g}$ of fr. wt.) (Table – 3). The chlorophyll content of green alga *E. intestinalis* was equal to green algae *Caulerpa scalpeliformis* and less than red alga *Acanthafera spicifera*^{32,33}. Ten estimated that the maximum carotenoid content in the brown seaweed was higher than the selected species of *Enteromorpha*.

4. CONCLUSION

The phytochemicals, biochemical and photosynthetic pigments were examined with four species of *Enteromorpha* which were collected from the intertidal coastal regions of Ramanathapuram District of Tamil Nadu, India. Phytochemicals were screened in all four species. Quantitatively analyzed biochemical was significantly

7. REFERENCES

1. Arnon DI. Copper enzymes in the isolated chloroplast, polyphenol oxidase in *Beta vulgaris*. *Plant Physiol.* 1949; 2:1-15.
2. Dinesh G., M, Sekar and R. Kannan Nutritive properties of seaweeds of Gulf of Mannar, Tamil Nadu. *Seaweed Res Utiln.* 2007;29(1, 2):125-32.
3. Folch J, Lees M and Sloane Stanley GH. A simple method for isolating and purifying total lipids from animal tissues. *J Biol Chem.* 1956; 226:497-509.
4. Horborne, I.B. Phytochemical methods. New York: Chapman & Hall; 1973. p. 288. doi: 10.1007/978-94-009-5570-7.
5. Kaliaperumal N., IR, Ramalingam Kalimuthu S, Ezhilvalavan, R. Seasonal changes in growth, biochemical constitutions and phycocolloid of some marine algae of Mandapam coast. *Seaweed Res Utiln.* 2002;24(1):73-7.
6. Kirk ITO, Allen RL. Dependence of chloroplast pigments synthesis on protein synthetic effects on actilione. *Biochem Biphys Res Cann.* 1965; 27: 523-30.
7. Lima-Filho IVM, Carvalho AFFU, Freitas SM, Melo VMM. Antibacterial activity of extracts of six macroalgae from the northeastern Brazilian coast. *Braz J Microbiol.* 2002;33(4):311-3. doi: 10.1590/S1517-83822002000400006.
8. Wells ML, Potin P, Craigie JS, Raven JA, Merchant SS, Helliwell KE, et al. Algae as nutritional and functional food sources: revisiting our understanding. *J Appl. Phycol.* 2017;29(2):949-82. doi: 10.1007/s10811-016-0974-5, PMID 28458464.
9. Miller JDA. Fats and steroids, (in), Physiology and Biochemistry of algae. New York: Academic Press. p. 929; 1962 Lewin RA, editor. Available from: <https://www.sciencedirect.com/science/article/pii/004313549090013V> [cited 15/2/2023].
10. Muthuraman, B and R, Ranganathan. Biochemical studies of some green algae of the Kanyakumari coast. *Seaweed Res Utiln.* 2004;26(1&2): 69-71.
11. Chudasama NA, Sequeira RA, Moradiya K and Prasad K. Seaweed Polysaccharide Based Products and Materials: an Assessment on Their Production from a Sustainability Point of View. *Molecules.* 2021;26(9):1.19. doi: 10.3390/molecules26092608, PMID 33947023.
12. Raymont J, EG, Austin and E. Lineford Biochemical studies on zooplankton. *The Biochem Compos Neomycis Integer J Cans Perm Empor Mer.* 1964; 28:354-63.
13. Peñalver, R, Lorenzo, JM., Ros G, Amarowicz R, Pateiro M and G. Nieto Seaweeds as a functional ingredient for a healthy diet. *Mar Drugs.* 2020;18(6):301. doi: 10.3390/MD18060301, PMID 32517092.
14. Roe JH. The determination of sugar in the blood and spinal fluid with anthrone reagent. *J Biol Chem.* 1955;212(1):335-43. doi: 10.1016/S0021-9258(18)71120-4. PMID 13233235.
15. Lomartire, S and AMM. Goncalves An overview of potential seaweed-derived bioactive compounds for pharmaceutical applications. *Mar Drugs.* 2022;20(2):141. doi: 10.3390/MD20020141, PMID 35200670.
16. Paul S, and R. Kundu Antiproliferative activity of methanolic extracts from two green algae, *Enteromorpha intestinalis* and *Rizocloniumriparium* on HeLa cells. *Daru.* 2013;21(1):72. doi: 10.1186/2008-2231-21-72, PMID 24355313.
17. Vasanthi HR, Jaswanth A, Krishnaraj V, Rajamanickam GV and A. Saraswathy. In vitro snake venom detoxifying action of some marine algae of Gulf of Mannar, southeast of India. *Phytother Res.* 2003;17(10):1217-9. doi: 10.1002/ptr.1342, PMID 14669260.
18. Wang X, Chen Y, Wang I, Liu Z and S Zhao. Antitumor activity of a sulfated polysaccharide from *Enteromorpha intestinalis* targeted against hepatoma through a mitochondrial pathway. *Tumour Biol.* 2014;35(2):1641-7. doi: 10.1007/s13277-013-1226-9. PMID 24197975.
19. Peñalver R, Lorenzo JM, Ros G, Amarowicz R, Pateiro M and G Nieto. Seaweeds as a functional ingredient for a healthy diet. *Mar Drugs.* 2020 Jun 5;18(6):301. doi: 10.3390/MD18060301, PMID 32517092.
20. Qin Y. Seaweed hydrocolloids as a thickening, gelling, and emulsifying agent in functional food products.

distributed among the species. In addition, the photosynthetic pigments were quantified in all the species. The present study shows that marine macro algae, like *Enteromorpha compressa*, *E. flexuosa*, *E. intestinalis*, and *E. prolifera* are high in nutritive properties. In addition, the green algae *Enteromorpha* species observed a high lipid value. The present findings will be useful for future bio-product productions such as cosmetics, skincare products, industrial food products, and pharmaceutical industries and also very useful to feed cattle and birds.

5. AUTHOR CONTRIBUTION STATEMENT

Sona performed all of the tasks necessary for this study and the publication of this manuscript.

6. CONFLICT OF INTEREST

Conflict of interest declared none.

Bioactive seaweeds for food applications 2018 Jan 1 (pp. 135-52). Academic Press.

21. Cofrades S, López-Lopez I, Bravo L, Ruiz-Capillas C, Bastida S and MT Larrea. Nutritional and antioxidant properties of different brown and red Spanish edible seaweeds. *Food Sci Technol Int.* 2010 Oct;16(5):361-70. doi: 10.1177/1082013210367049, PMID 21339154.

22. Roohinejad S, Koubaa M, Barba FI, Saljoughian S, Amid M and R. Greiner Application of seaweeds to develop new food products with enhanced shelf-life, quality and health-related beneficial properties. *Food Res Int.* 2017 Sep 1;99(3):1066-83. doi: 10.1016/j.foodres.2016.08.016, PMID 28865618.

23. Lange KW, Hauser I, Nakamura Y, Kanaya S. Dietary seaweeds and obesity. *Food Sci Hum Wellness.* 2015 Sep 1;4(3):87-96. doi: 10.1016/j.fshw.2015.08.001.

24. Sona P, Ravi P, Ambiga K, Manivannan M, Ashwathaman S, Subramanian G. Studies on phytochemicals of *Enteromorpha compressa* (L.) NEES-A green alga from Rameshwaram COASTLINE. TAMIL NADU, INDIA.

25. Yan X, Yang C, Lin G, Chen Y, Miao S, and B. Liu. Antidiabetic potential of green seaweed *Enteromorpha prolifera* flavonoids regulating insulin signaling pathway and gut microbiota in type 2 diabetic mice. *J Food Sci.* 2019 Jan;84(1):165-73. doi: 10.1111/1750-3841.14415, PMID 30569533.

26. Farasat M, Khavari-Nejad RA, Nabavi SM and F. Namjooyan. Antioxidant activity, total phenolics and flavonoid contents of some edible green seaweeds from northern coasts of the Persian Gulf. *Iran J Pharm Res.* 2014;13(1):163-70. PMID 24734068.

27. Yan X, Yang C, Lin G, Chen Y, Miao S and B Liu. Antidiabetic potential of green seaweed *Enteromorpha prolifera* flavonoids regulating insulin signaling pathway and gut microbiota in type 2 diabetic mice. *J Food Sci.* 2019 Jan;84(1):165-73. doi: 10.1111/1750-3841.14415, PMID 30569533.

28. Ganesan K, Suresh Kumar K, Subba Rao PV, Tsukui Y, Bhaskar N and M Hosokawa. Studies on the chemical composition of three species of *Enteromorpha*. *Biomed Prev Nutr.* 2014 Jul 1;4(3):365-9. doi: 10.1016/j.bionut.2014.04.001.

29. Shanab SM, Shalaby EA and El-Fayoumy EA. *Enteromorpha compressa* exhibits potent antioxidant activity. *J Biomed Biotechnol.* 2011 Jun; 2011: 726405. doi: 10.1155/2011/726405, PMID 21869863.

30. Chen K, Ríos JJ, Pérez-Gálvez A, Roca M. Comprehensive chlorophyll composition in the main edible seaweeds. *Food Chem.* 2017 Aug 1; 228:625-33. doi: 10.1016/j.foodchem.2017.02.036, PMID 28317773.

31. Lin A, Wang C, Qiao H, Pan G, Wang G and L. Song. Study the photosynthetic performances of *Enteromorpha prolifera* collected from the surface and bottom of the sea of Qingdao sea area. *Sci Bull.* 2009 Feb;54(3):399-404. doi: 10.1007/s11434-009-0025-6.

32. Shafique S, Siddiqui PI, Aziz RA, Burhan Z and SN Mansoor. Weight loss and changes in organic, inorganic, and chlorophyll contents in three species of seaweeds during decomposition. *Pak J Bot.* 2010 Aug 1;42(4):2599-604.

33. Bouhadi M, Cherifi O, Bahammou N, Cherifi K, Taibi M, Elkouali M. The effect of *Enteromorpha intestinalis* and *Corallina elongata* on physiological parameters of *Zea mays* L. *Arab Gulf J Sci Res.* 2022:303-13. doi: 10.51758/AGJSR-04-2021-0032.

34. Leskinen E, ALSTRÖM-RAPAPORT C and P. Pamilo. Phylogeographical structure, distribution and genetic variation of the green algae *Ulva intestinalis* and *U. compressa* (Chlorophyta) in the Baltic Sea area. *Mol Ecol.* 2004 Aug;13(8):2257-65. doi: 10.1111/j.1365-294X.2004.02219.x, PMID 15245399.

35. Blomster J, Maggs CA, Stanhope MJ. Molecular and morphological analysis of *Enteromorpha intestinalis* and *E. compressa* (Chlorophyta) in the British Isles. *J Phycol.* 1998 Apr;34(2):319-40. doi: 10.1046/j.1529-8817.1998.340319.x.

36. Cho M, Lee HS, Kang II, Won MH and S. You. Antioxidant properties of extract and fractions from *Enteromorpha prolifera*, a type of green seaweed. *Food Chem.* 2011 Aug 1;127(3):999-1006. doi: 10.1016/j.foodchem.2011.01.072, PMID 25214089.

37. Zhao S, He Y, Wang C, Assani I, Hou P and Y Feng. Isolation, characterization and bioactive properties of alkali-extracted polysaccharides from *Enteromorpha prolifera*. *Mar Drugs.* 2020 Nov 6;18(11):552. doi: 10.3390/md18110552, PMID 33172133.

38. Banu KS and L. Cathrine. General techniques involved in phytochemical analysis. *Int J Adv Res Chem Sci.* 2015 Apr;2(4):25-32.