



## Formulation and Evaluation of Captopril Loaded Niosomal Transdermal Films

Angilicam Avinash<sup>\*1, 2</sup> , P. Dwarakanadha Reddy<sup>3</sup> and S. V. Satyanarayana<sup>4</sup>

<sup>1</sup>Research Scholar, Research & Development, JNTUA, Ananthapuramu, Andhra Pradesh, India-515002

<sup>2</sup>Department of Pharmaceutics, Narayana Pharmacy College, Nellore, Andhra Pradesh, India-524003

<sup>3</sup>Department of Pharmaceutics, Annamacharya College of Pharmacy, Rajampet, Andhra Pradesh, India-516126

<sup>4</sup>Principal, JNTUA College of Engineering, Kalikiri, Annamayya Dist, Andhra Pradesh, India- 517234.

**Abstract:** Captopril was the first angiotensin-converting enzyme (ACE) inhibitor used for the management of hypertension. The aim of this study was to prepare and evaluate niosomal-loaded captopril transdermal films. Captopril has good solubility but has poor permeability and reduced bioavailability in the presence of food. The aim and objective are to improve bioavailability and permeability. The captopril-loaded niosomal formulations were prepared by thin film hydration technique, using materials like non-ionic surfactants such as Spans of different grades 20, 40, 60 and 80 and solvents like ethanol and chloroform. The FT-IR results revealed that there was no interaction between excipients and captopril. All the formulations showed better encapsulation efficiency. The dissolution studies showed prolonged drug release in comparison to pure captopril. On comprising all formulations, F3 showed sustained release of 98.44% up to 12hrs. The optimized niosomes of captopril were used to prepare transdermal films using methyl cellulose, HPMC E5, HPMC K4M and HPMC K15M as a film forming agents and dibutyl phthalate as a plasticizer. All the formulated captopril transdermal films were evaluated for drug content, folding endurance, weight variation and *in-vitro* drug permeation. The *in-vitro* drug permeation was found to be 99.58% over a period of 12 hrs. Based on the above results, administering niosomal-loaded captopril through the transdermal route is a better approach.

**Keywords:** Bioavailability, Captopril, Niosomes, Non-Ionic Surfactants, Permeability and Transdermal Films.

---

### \*Corresponding Author

Angilicam Avinash, Research Scholar, Research & Development, JNTUA, Ananthapuramu, Andhra Pradesh, India-515002; Department of Pharmaceutics, Narayana Pharmacy College, Nellore, Andhra Pradesh, India-524003



Received On 19 September, 2022

Revised On 28 November, 2022

Accepted On 2 December, 2022

Published On 2 January, 2023

---

### Funding

This research did not receive any specific grant from any funding agencies in the public, commercial or not for profit sectors.

### Citation

Angilicam Avinash, P. Dwarakanadha Reddy and S. V. Satyanarayana, Formulation and Evaluation of Captopril Loaded Niosomal Transdermal Films.(2023).Int. J. Life Sci. Pharma Res.13(1), P143-155 <http://dx.doi.org/10.22376/ijlpr.2023.13.1.P143-155>

This article is under the CC BY- NC-ND Licence (<https://creativecommons.org/licenses/by-nc-nd/4.0>)

Copyright © International Journal of Life Science and Pharma Research, available at [www.ijlpr.com](http://www.ijlpr.com)

Int J Life Sci Pharma Res., Volume13., No 1 (January) 2023, pp P143-155



## I. INTRODUCTION

The treatment of a chronic disease or an acute disease has been generally accomplished by delivery of drugs to patients through various conventional dosage forms like capsules, tablets, liquids, aerosols and parenteral. This category of drug delivery is well-known to afford a prompt drug release. So, to achieve and maintain the concentration of a drug within the pharmacologically effective range, it is often required to take this type of conventional dosage form many times a day. This results in fluctuations in plasma drug concentration. Continuous I.V. infusion provides better systemic drug delivery that can maintain a constant and prolonged drug levels within a concentration range, which is therapeutically effective for treatment. But it requires continuous hospitalization during treatment and requires medical supervision. New techniques are developed that can control the drug delivery rate, prolong the duration of drug activity, and target the drug delivery to a particular site. The idea of delivering a drug through the skin laid efforts in the pharmaceutical field to develop a Transdermal drug delivery system to treat angina, hypertension, motion sickness and hormone deficiency.<sup>1</sup> Transdermal drug delivery system provides advantages. For instance, it is painless and easy to administer, it protects the active drug moiety from gastric substances and it, avoids first-pass metabolism, improved patient compliance, controls absorption rate, interference due to the presence of food is minimized, suitable for unconscious patients and enables termination of drug delivery if any side effects observed.<sup>2</sup> According to a recent report, the value of the worldwide market for transdermal drug delivery was \$12.7 billion in 2005 and \$ 52.4 billion in 2020 and is expected to increase to \$87.3 billion by 2030. Hypertension is a major situation that affects a large population of the world. Currently, a large number of oral medications are used for the treatment.<sup>3</sup> Transdermal drug delivery is the better alternative approach in the case when oral drug delivery is contraindicated, or poor absorption of the drug from G.I. tract.<sup>4</sup> Captopril was the first angiotensin-converting enzyme (ACE) inhibitor widely used for the management of congestive heart failure and hypertension. Captopril is considered a drug of choice in antihypertensive treatment due to its low toxicity and effectiveness. It has a  $t_{1/2}$  of 2-3 hrs, but the action duration lasts 6-12 hrs. Captopril shows 75% bioavailability but reduces oral absorption to 30-50% in the presence of food, aiming to improve the bioavailability.<sup>5</sup> The reduced bioavailability in the presence of food and short half-life makes captopril a good candidate for a transdermal drug delivery system (TDDS). A TDDS containing captopril avoids the reduction of bioavailability by drug intake or concomitant food and provides continuous dosing of the drug with improved patient compliance.<sup>6</sup> The captopril oxidation rate in dermal homogenate is significantly lower than the intestinal homogenate because the captopril disulfide, an oxidative product of captopril shows poor absorption from the intestine.<sup>7</sup> When administered initially, captopril causes hypotension, which can harm congestive heart failure and diuretic-treated patients. Persistent hypotension may cause some trouble in myocardial infarction patients.<sup>8</sup> Therefore, using a transdermal drug delivery system of captopril can reduce the side effects. Niosome carriers, known for their potential in topical drug delivery, have been used to transport captopril molecules in the skin layer.<sup>9</sup> Niosomes of

Captopril represents possible sustained-release formulation that increases the duration and magnitude of captopril<sup>10</sup>. The development of a once-daily captopril formulation would be a significant advantage for patient compliance, accompanied by minimizing drug side effects due to the reduction of drug blood concentration fluctuations in long-term therapy<sup>11, 12</sup>. It is well-known that transdermal applications greatly benefit from protecting drugs from the hepatic first-pass effect. However, the stratum corneum layer of the skin forms a barrier, resulting in slow absorption at the application site<sup>13</sup>. Captopril is generally marketed in the conventional dosage form of a tablet, usually with a strength of 12.5–50 mg. When the oral route administers the drug, it undergoes first-pass hepatic metabolism. The conventional tablet and capsule are administered 3 or 4 times a day due to their short biological half-life of about two hours. The bioavailability of captopril is reduced to 50% in the presence of food. The sustained release forms are administered two times a day due to their limited residence time in the gastrointestinal tract. These limitations of captopril in a conventional dosage form can be overcome by administering captopril through other routes of administration<sup>14</sup>. The main objective of the present research work was to develop transdermal films of captopril that deliver the captopril at a controlled rate and evaluate the *in-vitro* characteristics of the captopril transdermal films. The research work's novelty is formulating the transdermal films using captopril-loaded niosomes instead of pure captopril, as Niosomal-loaded captopril showed enhanced drug release<sup>15</sup>. Niosomal-loaded captopril transdermal films were developed and examined due to their higher delivery rate, minimum film area, and low drug concentration.<sup>16</sup>

## 2. MATERIALS AND METHODS

### 2.1. Materials

Captopril (A to Z Pharmaceuticals Ltd., Ambattur), MC, HPMC E5, HPMC K4M and HPMC K15M Di butyl phthalate (excel organic Pvt. Ltd., Chennai), Acetone (Merk Private Ltd., Mumbai), Ethanol (Changshu Hongsheng Fine Chemical Co. Ltd, China).

### 3. PREPARATION OF NIOSOMAL-LOADED CAPTOPRIL TRANSDERMAL FILMS

Niosomes, previously loaded with captopril by thin film hydration method, was used in the films' preparation. Transdermal films containing captopril were prepared using different concentrations of polymers (Methylcellulose, HPMC E5, HPMC K4M and HPMC K15M) individually or in combination. The drug concentration should be kept constant. The required amount of niosomal captopril (25 mg) and polymers were dispersed in solvents (Acetone and ethanol) and allowed to stir until the drug's and polymer's complete solubilization. After uniform mixing of the drug and polymer, the solution was allowed to stand for 10-20 minutes to remove air bubbles, and the final solution was poured into a petri dish. The polymeric drug solution allowed for evaporating for 24hrs to form a matrix film. After drying, the films were taken from the petri dish, cut to 2cm×2 cm, wrapped in butter paper and stored in desiccators for further studies.<sup>17, 18</sup> The compositions of different formulations are shown in Table I and 2.

**Table 1: Composition of captopril transdermal films**

Quantities (mg)	F1	F2	F3	F4	F5	F6	F7	F8	F9	F10
Captopril	240.62	240.62	240.62	240.62	240.62	240.62	240.62	240.62	240.62	240.62
Methylcellulose	200	-	-	-	100	100	100	-	-	-
HPMC E5	-	200	-	-	100	-	-	100	100	-
HPMC K4M	-	-	200	-	-	100	-	100	-	100
HPMC K15M	-	-	-	200	-	-	100	-	100	100
Di butyl phthalate (ml)	1	1	1	1	1	1	1	1	1	1
Acetone (ml)	3	3	3	3	3	3	3	3	3	3
Ethanol (ml)	6	6	6	6	6	6	6	6	6	6

4 cm<sup>2</sup> contains 25 mg captopril  
 38.5 cm<sup>2</sup> contains 240.62 mg of captopril  
 240.62 mg captopril is equivalent to 1.20 g of niosomal-loaded captopril

**Table 2: Composition of Niosomal Loaded Captopril Transdermal Films**

Formulation code	Plasticizer % (Di butyl phthalate)	Polymer %				Penetration enhancer % (Ethanol)
		MC	HPMC E5	HPMC K4M	HPMC K15M	
F1	5	100	-	-	-	10
F2	5	-	100	-	-	10
F3	5	-	-	100	-	10
F4	5	-	-	-	100	10
F5	5	50	50	-	-	10
F6	5	50	-	50	-	10
F7	5	50	-	-	50	10
F8	5	-	50	50	-	10
F9	5	-	50	-	50	10
F10	5	-	-	50	50	10

#### 4. EVALUATION OF NIOSOMAL LOADED CAPTOPRIL TRANSDERMAL FILMS

##### 4.1. Compatibility study using FTIR and DSC

Bruker company FT-IR was used to determine the pure drug's spectrums (Captopril) and its physical mixtures with polymers using the KBr pelletization method to find any possible reactions between the polymers and captopril. DSC Q20 V24.11 Build 124 was used to determine the potential chemical or physical interaction between polymers and captopril.<sup>19, 20</sup>

##### 4.2. The physical appearance of the Formulation

The prepared films were physically examined for colour, transparency and surface texture.<sup>21</sup>

##### 4.3. Thickness

The thickness of the captopril-loaded films was measured at

different points using a digital micrometre and determine the average thickness to ensure the uniform thickness of the film.<sup>22, 23</sup>

##### 4.4. Weight Uniformity

Five different films from individual formulations were weighed using high-accuracy digital balance, and the average weight was calculated. The individual weight should not deviate extensively from the average weight.<sup>24, 25</sup>

##### 4.5. Percentage of moisture loss

The prepared films were weighed individually and stored in a desiccator containing calcium chloride at room temperature for 24hrs. After 24hrs, the films were reweighed, and the percentage moisture content was determined using the following equation.<sup>26</sup>

$$\% \text{ Moisture loss} = \frac{\text{Initial weight} - \text{Final weight}}{\text{Final weight}} \times 100$$

##### 4.6. Percentage moisture uptake

The weighed films were kept in desiccators at room temperature for 24hrs containing a saturated potassium chloride solution. After 24hrs, the films were reweighed, and the percentage moisture uptake was determined using the following formula.<sup>27</sup>

$$\% \text{ Moisture Uptake} = \frac{\text{Final weight} - \text{Initial weight}}{\text{Initial weight}} \times 100$$

#### 4.7. Folding endurance

The folding endurance was measured manually for the formulated films. A film of 2×2 cm was taken and folded at the same place until it breaks. The number of times the film would be folded at the same position without breaking gives the value of folding endurance.<sup>28, 29</sup>

#### 4.8. Surface pH

Transdermal films were left to swell for 1 hr on the surface of the agar plate prepared by dissolving 1% (w/v) agar in warmed phosphate buffer of pH 7.4 with continuous stirring and then poured the solution into the glass dish and allowed to stand till it forms a gel at room temperature. The surface pH was measured using a pH meter placed on the surface of the swelled film.<sup>30</sup>

#### 4.9. Determination of tensile strength

The tensile strength is determined as the stretching force applied to the film at which it breaks. A specified weight was hung from the film through the sample such that a pulling force was produced. The force applied was measured in g/cm<sup>2</sup> on the load cell of the apparatus.<sup>31</sup>

#### 4.10. Drug content

A film of 4 cm<sup>2</sup> was placed in a standard flask containing 10 ml of phosphate buffer of pH 7.4 and was sonicated for 20 mins using an ultrasonicator. The volume was made up to 100 ml, and the absorbance was measured at 212 nm by Spectrophotometer with suitable dilutions.<sup>32, 33</sup>

#### 4.11. In-vitro permeation study

An *in-vitro* permeation study of prepared Captopril films was carried out using a Franz diffusion cell with 60ml capacity. Dialysis membrane (Hi-media) was used as a diffusion

membrane. The diffusion cell was filled with phosphate buffer pH 7.4; the dialysis membrane was placed on the cell. The temperature was maintained at 37 ± 0.5°C. At regular time intervals, 5ml samples were withdrawn and replaced with phosphate buffer pH 7.4 after each sampling for 12hrs. The samples drawn were filtered and analyzed. The amount of permeated drug was determined using a U.V. Spectrophotometer at 212 nm. The experiments were done in triplicate. The amount of drug diffused was calculated for all formulations.<sup>5, 34</sup>

#### 4.12. Stability Analysis

The optimized formulation transdermal films were stored at 40°C ± 2°C and 75% ±5% R.H. in stability chambers for three months. After three months, films were evaluated for weight variation, thickness and drug content.<sup>35</sup>

### 5. RESULTS AND DISCUSSION

Prepared films were found to be smooth, flexible and homogeneous<sup>21</sup>. The prepared formulations were evaluated for different physicochemical properties, as shown in Table 3. The weight of the films varied from 141.33 to 174.33 mg. The thickness of the films ranged from 222.67 to 274 µm<sup>7</sup>. The thickness of the films was uniform, indicating the uniform distribution of drug and polymer solution. The low standard deviation values represent uniformity in all formulations<sup>36</sup>. The percentage of Moisture loss of the prepared transdermal films was found to be between 2.31 to 6.12. The rate of Moisture uptake of the prepared transdermal films was found to be between 1.46 to 7.18. All the formulations showed lower moisture content. Lower moisture content in the formulations helps them to remain stable and become a wholly dried and brittle film. Low moisture uptake protects the formulated films from microbial contamination and bulkiness<sup>37</sup>. The variation in moisture uptake depends on the polymer's ability to absorb the moisture<sup>38</sup>.

**Table 3: Different Physico-Chemical Properties of Captopril Films**

Formulation code	Thickness (µm)	Weight Uniformity (mg)	% Moisture loss	% Moisture uptake
F1	254.67±2.52	163.00±1.00	3.01±0.79	2.75±0.79
F2	222.67±1.53	141.33±2.52	5.98±1.33	7.18±1.51
F3	234.00±1.00	146.67±2.08	6.12±1.07	5.56±2.40
F4	243.67±3.06	152.33±1.15	4.23±1.90	3.84±2.07
F5	236.33±0.58	148.00±1.00	5.36±3.18	3.67±1.48
F6	244.67±1.53	153.33±1.53	5.60±2.76	3.13±2.05
F7	274.00±2.65	174.33±3.06	2.31±1.27	1.46±0.64
F8	228.33±1.15	146.67±2.08	4.38±0.02	5.26±1.31
F9	240.67±1.53	149.33±1.53	3.59±2.48	4.98±1.49
F10	261.00±2.00	168.67±1.53	3.19±1.55	2.43±0.59

Values are mean ± S.D; (n= 3)

Drug content was found to be in the range of 85.99 to 98.51%. The drug content of formulation F7 was higher than other formulations. This shows that the drug dispersed uniformly throughout the polymeric film<sup>39,40</sup>. The folding endurance test was performed manually, and maximum folding endurance was observed in formulation F7. If the films

showed cracks on their surface, it was considered endpoint<sup>1</sup>. The results were satisfactory, indicating that the films would not break and would maintain their integrity when used<sup>41</sup>. The pH of all the formulations mimics the skin's pH of 4<sup>2</sup>. Tensile strength was in the range of 205 to 333.3. The results are shown in Table 4.

Formulation code	% Drug content	Folding endurance	Surface pH	Tensile Strength (g/cm <sup>2</sup> )
F1	95.63±0.10	97.3±2.5	5.40±0.17	235.3±3.2
F2	85.99±0.16	84.0±3.6	5.33±0.15	205.0±3.6
F3	89.80±0.18	92.7±1.5	5.43±0.06	222.3±4.0
F4	94.14±0.22	107.3±2.1	5.53±0.21	259.3±2.5
F5	87.89±0.24	87.7±1.5	5.60±0.26	210.3±4.0
F6	94.17±0.31	110.7±2.1	5.83±0.25	265.7±4.7
F7	98.51±0.52	138.0±3.6	5.80±0.46	333.3±4.5
F8	89.46±0.53	87.3±1.5	5.70±0.53	209.7±4.0
F9	88.35±0.55	90.7±2.1	5.77±0.42	218±4.6
F10	95.32±0.52	99.3±2.5	5.50±0.79	237.7±4.2

Values are mean ± S.D; (n= 3)

### 5.1. Compatibility study using FT-IR and DSC

Pure captopril showed principal absorption peaks at 672.35 cm<sup>-1</sup> (C-S stretch), 1193.64 cm<sup>-1</sup> (C-N stretch), 1467.12 cm<sup>-1</sup> (C-H bend), 1581.90 cm<sup>-1</sup> (N-H bend) and 1740.90 cm<sup>-1</sup> (C=O stretch). The identical peaks of C-S stretching, C-N stretching, C-H bending, N-H bending, and C=O stretching vibrations were also noticed in the spectra of the drug mixed with polymers. FT-IR spectra revealed no interaction

between the polymers and the drug used for transdermal film preparation<sup>20,43</sup>. The results are shown in Table No.5 and Figures 1-5. The DSC studies of captopril showed a peak at 110.41°C. The DSC analysis of physical mixtures of captopril showed little change in the melting point of captopril. DSC studies revealed the absence of interactions between the drug and polymers<sup>44</sup>. The results are shown in Figure 6-10

Table 5: FT-IR Interpretations of Pure Drug and Excipients

S. No	Functional group	Characteristic peaks	Observed peaks				
			Captopril (CAP)	CAP: MC	CAP: HPMC E5	CAP: HPMC K4M	CAP: HPMC K15M
1	C-S (Stretching)	600-800 cm <sup>-1</sup>	672.35 cm <sup>-1</sup>	670.09 cm <sup>-1</sup>	671.07 cm <sup>-1</sup>	668.40 cm <sup>-1</sup>	669.76 cm <sup>-1</sup>
2	C-N (Stretching)	1020-1250 cm <sup>-1</sup>	1193.64 cm <sup>-1</sup>	1220.93 cm <sup>-1</sup>	1197.79 cm <sup>-1</sup>	1220.85 cm <sup>-1</sup>	1223.39 cm <sup>-1</sup>
3	C-H (Bending)	1440-1480 cm <sup>-1</sup>	1467.12 cm <sup>-1</sup>	1476.40 cm <sup>-1</sup>	1464.85 cm <sup>-1</sup>	1463.76 cm <sup>-1</sup>	1447.18 cm <sup>-1</sup>
4	N-H (Bending)	1590-1650 cm <sup>-1</sup>	1581.90 cm <sup>-1</sup>	1581.63 cm <sup>-1</sup>	1584.28 cm <sup>-1</sup>	1581.78 cm <sup>-1</sup>	1581.51 cm <sup>-1</sup>
5	C=O (Stretching)	1690-1760 cm <sup>-1</sup>	1740.90 cm <sup>-1</sup>	1743.24 cm <sup>-1</sup>	1740.96 cm <sup>-1</sup>	1741.47 cm <sup>-1</sup>	1743.17 cm <sup>-1</sup>

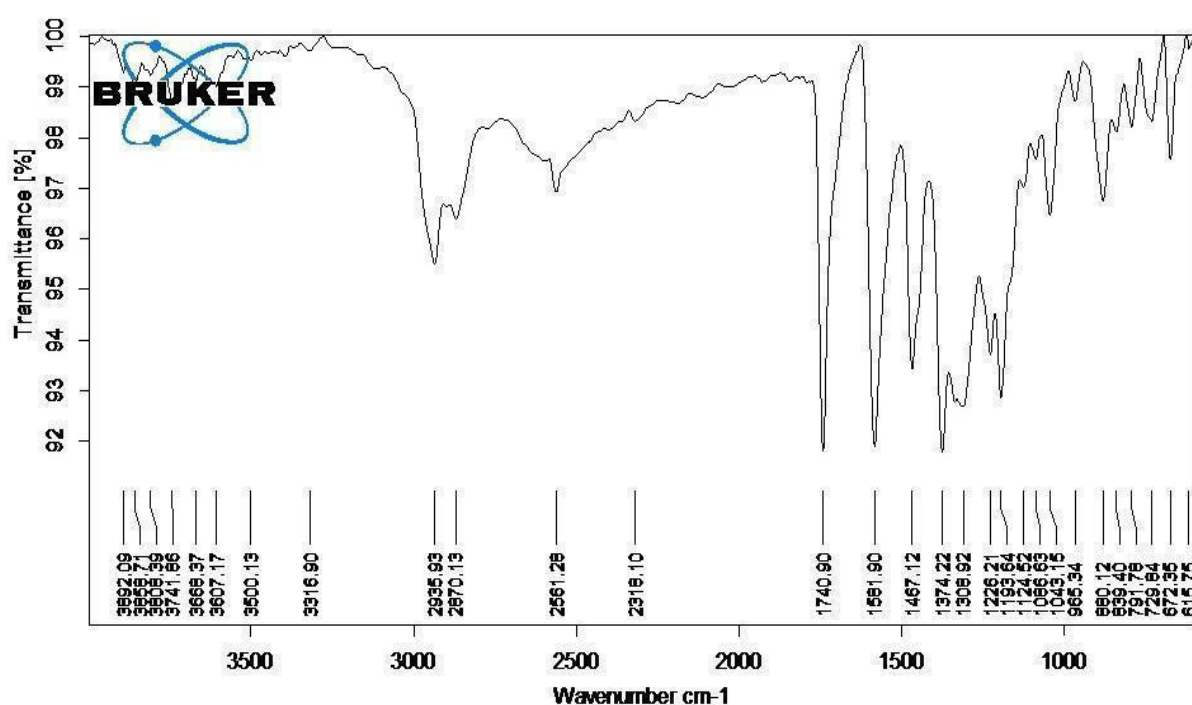


Fig 1: FT-IR Spectrum of captopril



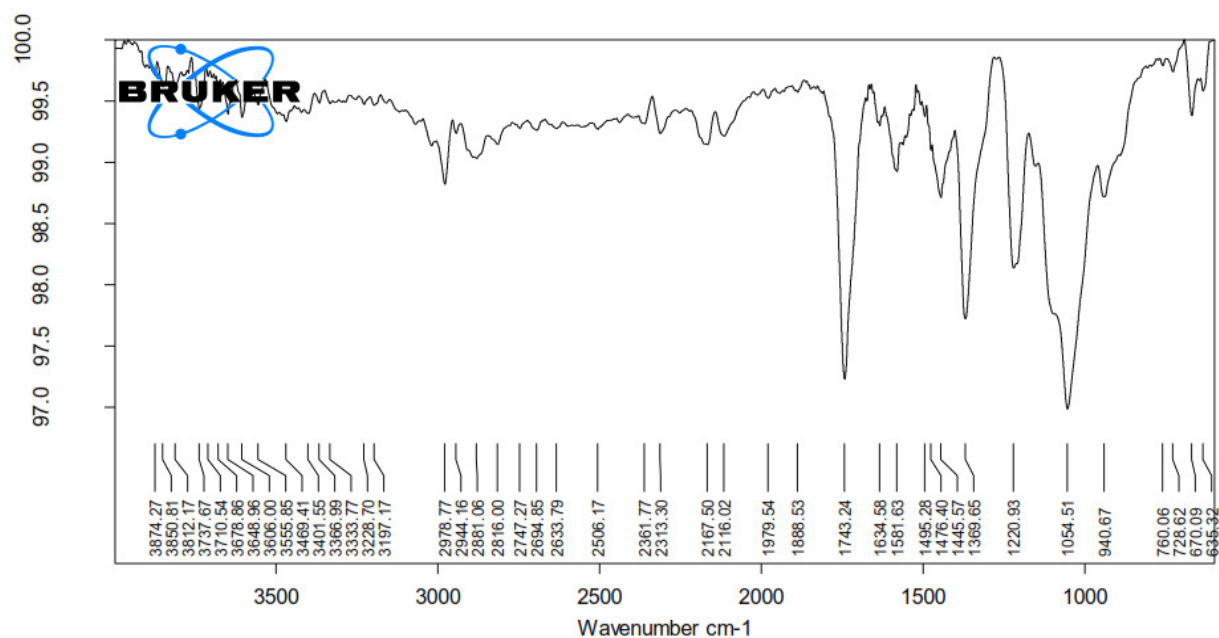


Fig 2: FT-IR Spectrum of captopril with methylcellulose

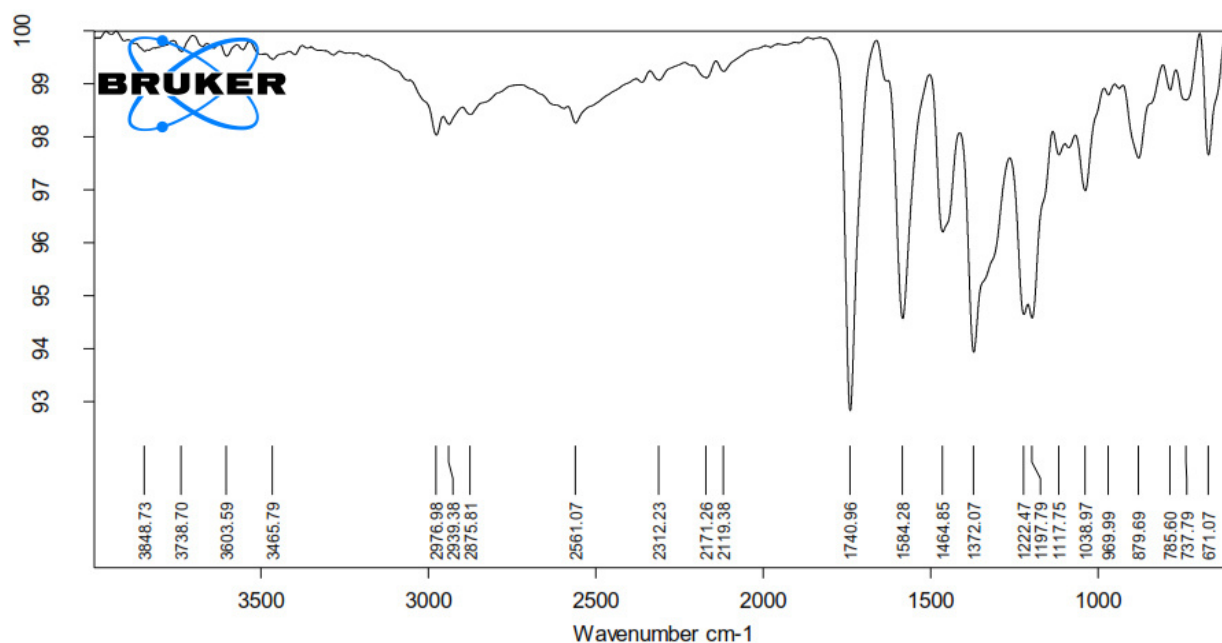


Fig 3: FT-IR Spectrum of captopril with HPMC E5

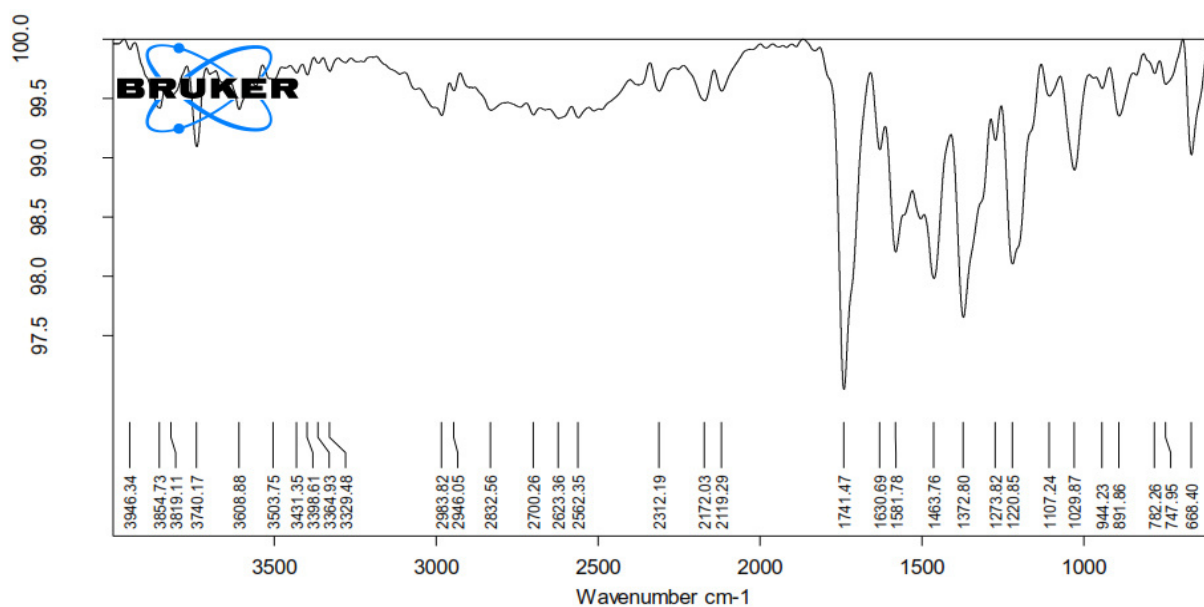


Fig 4: FT-IR Spectrum of captopril with HPMC K4M

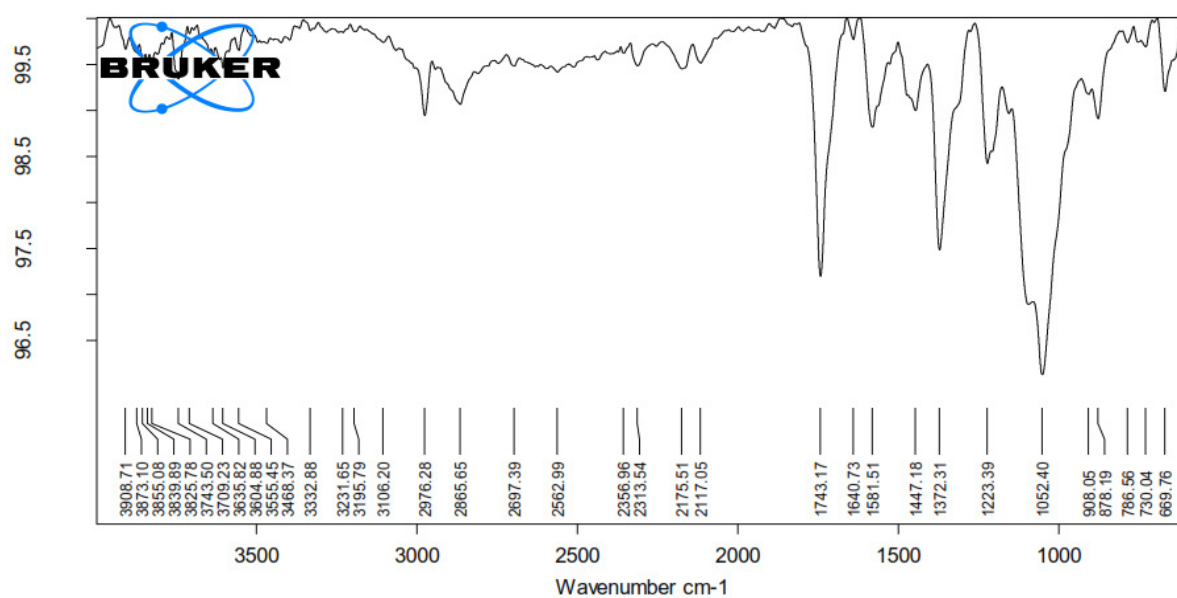


Fig 5: FT-IR Spectrum of captopril with HPMC K15M

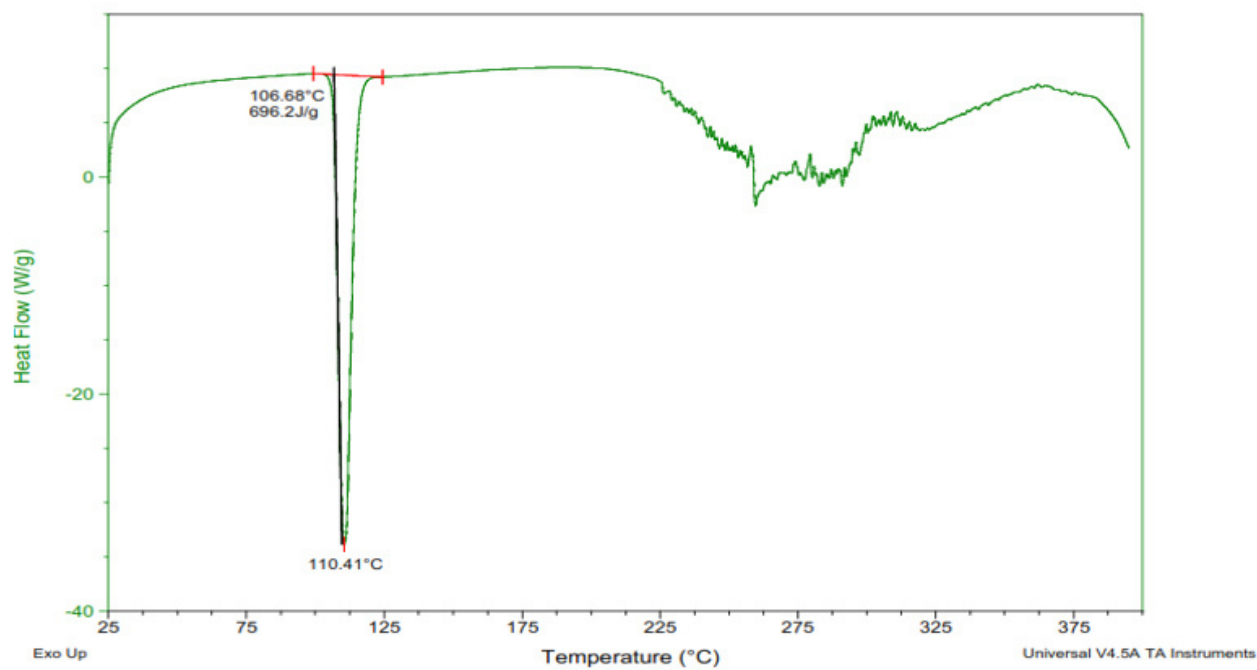


Fig 6: DSC thermogram of captopril

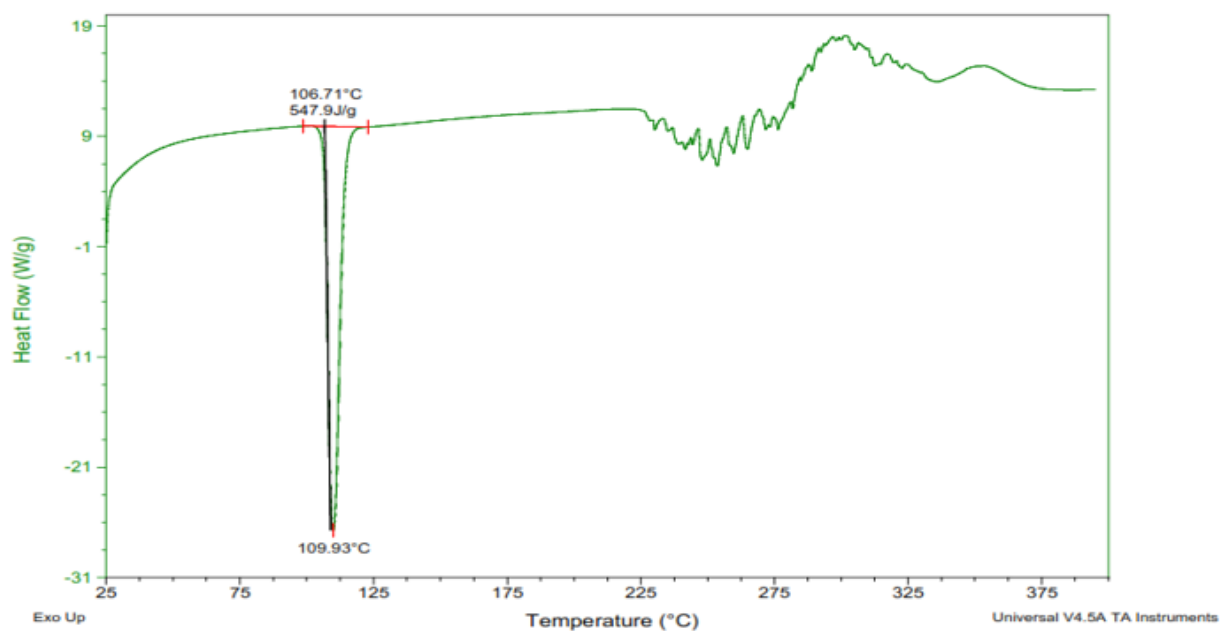
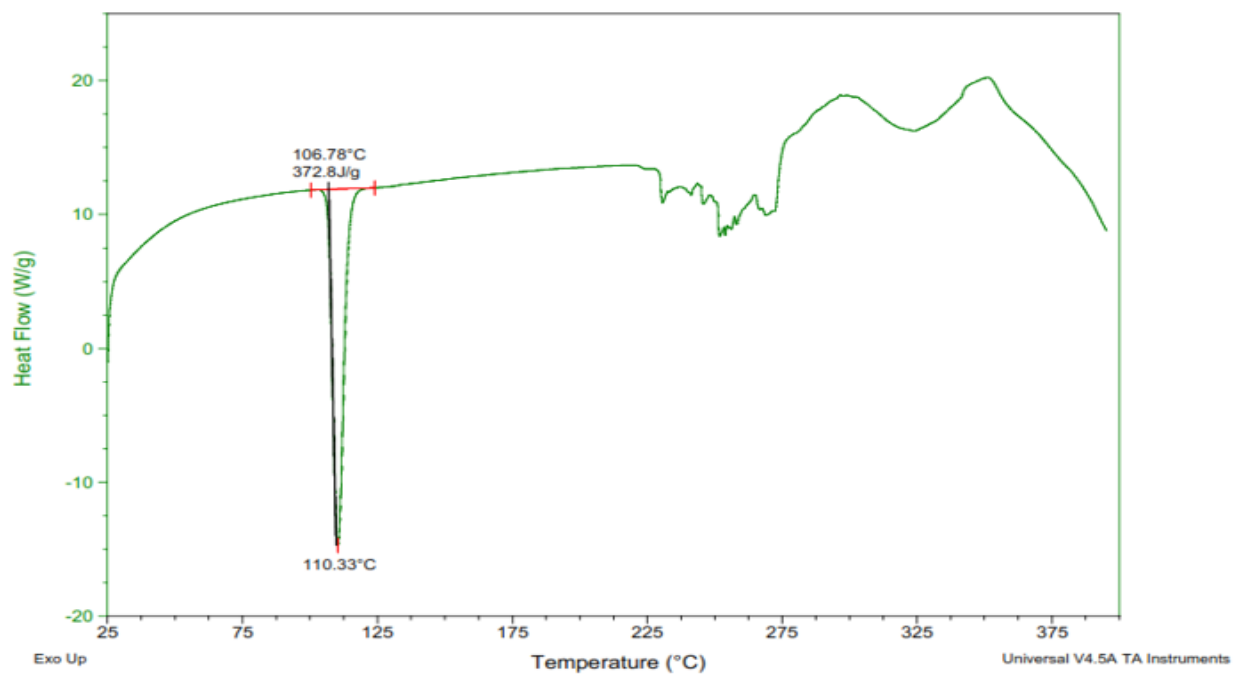
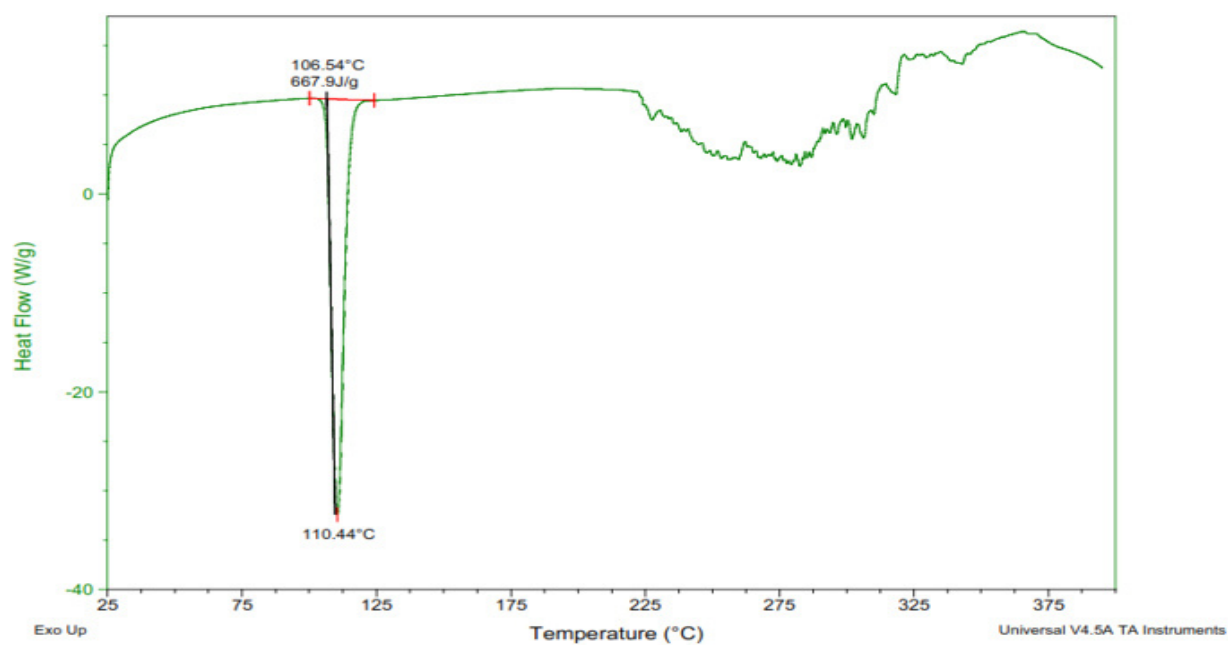


Fig 7: DSC thermogram of captopril with methylcellulose

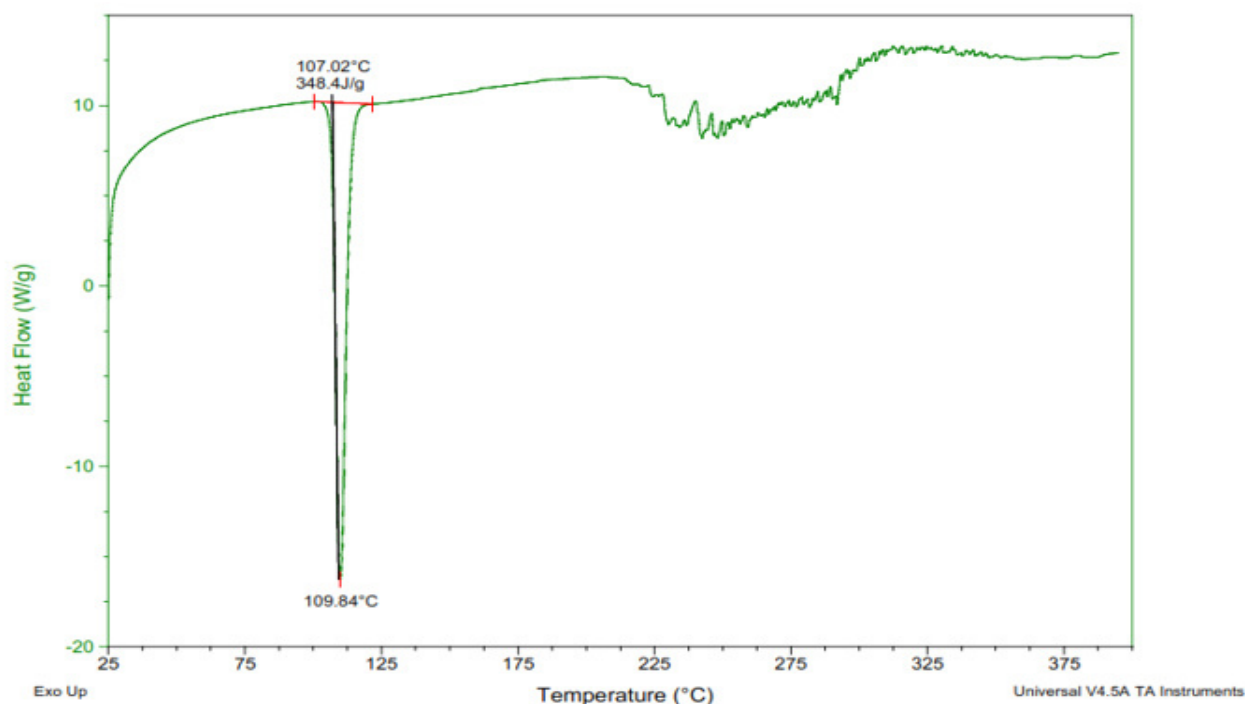




**Fig 8: DSC thermogram of captopril with HPMC E5**



**Fig 9: DSC thermogram of captopril with HPMC K4M**



**Fig 10: DSC thermogram of captopril with HPMC K15M**

## 5.2. *In-vitro* permeation study

Captopril transdermal films were prepared by the Solvent casting method. All the formulations viz. F1-F10 cumulative permeation data are shown in Figure.11. *In-vitro* drug permeation studies were carried out using a Franz diffusion cell using a Hi-media dialysis membrane. The result of

diffusion studies indicates that formulation F7 containing MC and HPMC K15M showed the highest drug permeation, 99.58 %, in 12 hrs. The high viscosity of MC and the swelling nature of HPMC may cause sustained release when compared to other formulations. The samples were analyzed spectrophotometrically at 212 nm<sup>45</sup>.

## 5.3. Stability Analysis

Table 6: Stability Analysis of optimized Formulation (F7)					
S. No	Observation	Initial	After 1 month	After 2 months	After 3 months
1	Average weight (g)	174.33±3.06	173.00±2.00	172.33± 1.15	172.67± 3.51
2	Thickness (µm)	274.00±2.65	273.33±2.08	270.33±1.53	267.67±2.08
3	Folding Endurance	138.0±3.61	137.0±2.65	136.0±2.00	134.33±3.51
4	Tensile Strength	333.3±4.51	331.0±3.00	328.0±3.00	314.33±4.16
5	Drug Content	98.51±0.52	98.44±0.11	98.23±0.11	98.13±0.65
6	Surface pH	5.80±0.46	5.80±0.50	5.77±0.42	5.77±0.35
7	Drug Permeation	99.58±0.31	99.20±0.12	99.06±0.36	98.96±0.54

Values are mean ± S.D; (n= 3)

The results from stability analysis indicated that there was no significant difference. In addition, stability studies showed that optimized formulation F7 was stable. Therefore, all the stability studies were conducted as per ICH guidelines.

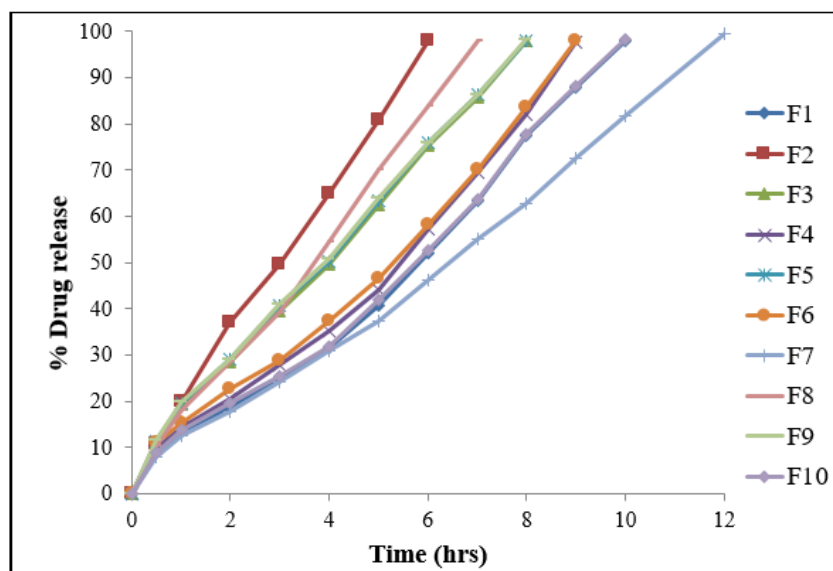


Fig 11: *In-vitro* drug permeation profile of captopril transdermal films

## 6. CONCLUSION

The optimized niosomal loaded captopril was taken for preparation of transdermal films by Solvent casting Method. MC, HPMC E5, HPMC K15M and HPMC K4M were used for the formulation of transdermal films. Dibutyl phthalate was used as a plasticizer, and ethanol was used as a permeation enhancer. Preformulation studies were performed to find drug-excipient interactions. FT-IR spectra revealed no interaction between the drug and the polymers used to prepare transdermal films. The prepared Captopril transdermal films were evaluated for thickness, weight uniformity, % Moisture loss, % Moisture uptake, Folding endurance, Tensile strength, Surface pH and % Drug content. *In addition, in-vitro* permeation studies were performed. Good results were obtained for evaluated parameters and *in vitro* studies. F7 formulation containing HPMC K4M and HPMC

K15M exhibited the release of 99.58% up to 12 hrs. The results from stability analysis indicated that there was no significant difference and found to be stable.

## 7. AUTHORS CONTRIBUTION STATEMENT

Mr Angilicam Avinash gathered the materials required and designed the formulation methodology for this research work. Dr P. Dwarakanadha Reddy and Prof. S. V. Satyanarayana performed formulations trials for designing the final methods. All authors discussed the results obtained and contributed to writing the manuscript.

## 8. CONFLICT OF INTEREST

Conflict of interest declared none.

## 9. REFERENCES

- Duraivel S., Rajalakshmi A.N and Debjit Bhowmik. Formulation and evaluation of captopril Transdermal Patches. *Elixir International Journal*.2014; 76: 28209-28213.  
[https://www.elixirpublishers.com/articles/1414820802\\_76%20\(2014\)%2028209-28213.pdf](https://www.elixirpublishers.com/articles/1414820802_76%20(2014)%2028209-28213.pdf)
- Md. Khaja, Muneer Syed, D. Srinivasa Rao. Design, Development and Evaluation of Transdermal Drug Delivery of Captopril, an Antihypertensive Drug. *Indo American Journal of Pharmaceutical Sciences*. 2014; 1(5): 305-314.  
<http://www.iajps.com/pdf/november/captopril.pdf>
- Darren R. Gullick, W. John Pugh, Matthew J. Ingram, Paul A. Cox and Gary P. Moss. Formulation and characterization of a captopril ethyl ester drug-in-adhesive-type patch for percutaneous absorption. *Drug Development and Industrial Pharmacy*. 2010; 36(8): 926–932.  
<https://doi.org/10.3109/03639040903585135>.
- Oya KERIMOGLU, Sevinç SAHBAZ, Özer SEHIRLI, Betül DORTUNÇ, Göksel SENER. Mechanical evaluation of matrix type transdermal therapeutic systems containing captopril. *Marmara Pharmaceutical Journal*. 2015; 19: 67-72.  
<https://dergipark.org.tr/en/download/article-file/166071>.
- Goswami D. S., Uppal N. Formulation and Evaluation of Transdermal Delivery System of an Antihypertensive Drug. *Journal of Applied Pharmaceutical Research*. 2013; 1(1): 31-35.  
<https://www.japtronline.com/index.php/joapr/article/view/1/5>.
- Eun-Seok Park, Seok-Jung Chang, Yun-Seok Rhee, and Sang-Cheol Chi. Effects of Adhesives and Permeation Enhancers on the Skin Permeation of Captopril. *Drug Development and Industrial Pharmacy*. 2001; 27(9): 975–980. DOI: 10.1081/ddc-100107679. PMID: 11763476.
- Zhou XH, Li Wan PA. Stability and in-vitro absorption of captopril, enalapril and lisinopril across the rat intestine. *Biochem Pharmacol*. 1994; 47: 1121-1126.
- Tripathi KD. *Essentials of Medical Pharmacology*. New Delhi, India, Jaypee Brothers, 2003 pp 449-454.
- Ankur Gupta, Sunil Kumar Prajapati, M Balamurugan, Mamta Singh, Daksh Bhatia. Design and Development of a Proniosomal Transdermal Drug Delivery System for Captopril. *Tropical Journal of*

- Pharmaceutical Research. 2007; 6 (2): 687-693. doi: 10.4314/tjpr.v6i2.2.
10. Mariangela de Burgos M de Azevedo, Ljubica Tasic, Juliana Fattori, Fábio hs rodrigues, Fabiana c cantos, Leandro P ribeiro, Vanice de Paula, Danielle lanzer and robson As santos. New formulation of an old drug in hypertension treatment: the sustained release of captopril from cyclodextrin nanoparticles. International Journal of Nanomedicine. 2011;6 1005–1016. DOI: 10.2147/IJN.S18999.
11. Wilding, I. R.; Davis, S. S.; Bkhshae, M.; Stevens, H. N.; Sparrow, R. A. Gastrointestinal transit and systemic absorption of captopril from a pulsed released formulation. *Pharm. Res.* 1992, 9 (5): 654-657.
12. Ankur Gupta, Sunil Kr. Prajapati, Mamta Singh, and M. Balamurugan. Proniosomal Powder of Captopril: Formulation and Evaluation. *Molecular Pharmaceutics*, 2007, 4(4): 596-599. DOI: 10.1021/mp0700110.
13. Soumya Singh. Niosomes: A Role in Targeted Drug Delivery System. *IJPSR*, 2013; Vol. 4(2): 550-557. DOI: [http://dx.doi.org/10.13040/IJPSR.0975-8232.4\(2\).550-57](http://dx.doi.org/10.13040/IJPSR.0975-8232.4(2).550-57).
14. Manish Kumar Gupta, Dr. Deepak Prakash, Dr. Brahmeshwar Mishra. Development and Characterization of Microparticulated Drug Delivery System of Captopril. *Indo American Journal of Pharm Research*.2013; 3(10):8323-8332.
15. Angilicam Avinash, P. Dwarakanadha Reddy, S. V. Satyanarayana. Design and Evaluation of Captopril-loaded Niosomes. *Asian Journal of Pharmaceutics*. 2022; 16 (3): 261-267. DOI: <https://doi.org/10.22377/ajp.v16i3.4475>.
16. Nimesh Goswami, Paresh Prajapati. Quality by Design (QBD) to Optimization of Semi-Solid Suspension Type of Captopril Transdermal Drug Delivery System. *International Journal of Research and Development in Pharmacy and Life Sciences*. 2016; 5(2): 2023-2038. <https://ubipayroll.com/IJRDPL/index.php/ijrdpl/article/view/215/217>.
17. Shefrin S., Sreelaxmi C. S., Vishnu Vijayan, Sreeja C. Nair. Anti-Epileptic Drug Loaded Niosomal Transdermal Patch for Enhanced Skin Permeation. *International Journal of Applied Pharmaceutics*. 2019; 11(2): 31-43. <https://doi.org/10.22159/ijap.2019v11i2.27034>
18. Vandana Mohabe, Rachna Akhand and Anupam Kumar Pathak. Preparation and Evaluation of Captopril Transdermal Patches. *Bulletin of Pharmaceutical Research*.2011; 1(2):47-52. [https://www.researchgate.net/publication/328213985\\_PREPARATION\\_AND\\_EVALUATION\\_OF\\_CAPTOPRIL\\_TRANSDERMAL\\_PATCHES](https://www.researchgate.net/publication/328213985_PREPARATION_AND_EVALUATION_OF_CAPTOPRIL_TRANSDERMAL_PATCHES).
19. Nukaraju K, Veerraju T and Satyavathi K, Formulation and Evaluation of Transdermal Patches of Cetrizine Dihydrochloride. *International Journal of Pharmaceutical Sciences Review and Research*. 2014; 25(1): 178-182. [https://nanopdf.com/download/5b16c92650534\\_pdf](https://nanopdf.com/download/5b16c92650534_pdf)
20. N. Swathi, D. Jayaprakash. Formulation Development and Evaluation of Captopril Mouth Dissolving Films. *International Journal of ChemTech Research*. 2019; 12(03): 17-27.
21. PravinUttekar, Akshata Kulkarni, Pravin Chaudhari, Manoj Dhage and Vishal Dhangarmali. Formulation and Evaluation of Captopril Transdermal patches for the treatment of hypertension. *Der Pharmacia Lettre*. 2016; 8 (5):12-16. <https://www.scholarsresearchlibrary.com/articles/formulation-and-evaluation-of-captopril-transdermal-patches-for-the-treatment-of-hypertension.pdf>
22. Vijayan V., Sumanth M.H., Suman L., Vinay T., SrinivasraoD., Jayaraj Kumar K, Development and Physiochemical, *In-Vitro* Evaluation of Antihypertensive Transdermal Patches, *Journal of Pharmaceutical Sciences and Research*, 2010, 2(3),171-177. <https://citeseerx.ist.psu.edu/viewdoc/download?doi=10.1.1.295.1329&rep=rep1&type=pdf>
23. Gajanan Darwhekar, Dinesh Kumar Jain, Vinod Kumar Patidar. Formulation and Evaluation of Transdermal Drug Delivery System of Clopidogrel Bisulfate. *Asian Journal of Pharmacy and Life Science*. 2011; 1(3):269-278. <https://www.yumpu.com/en/document/view/31127487/formulation-and-evaluation-of-transdermal-drug-delivery-system-of>
24. NavneetVerma, ShikhaDeshwal, Design and *In Vitro* Evaluation of Transdermal Patches Containing Ketoprofen, *World Journal of Pharmaceutical Research*, 2014, 3(3), 3930-3944. [https://wjpr.s3.amazonaws.com/article\\_issue/1398843102.pdf](https://wjpr.s3.amazonaws.com/article_issue/1398843102.pdf)
25. Anisree G.S, Ramasamy C, John Wesley.I, Bincy Mary Koshy. Formulation of Transdermal Drug Delivery System of Metoprolol Tartrate and its Evaluation. *Journal Pharmaceutical Sciences & Research*. 2012; 4(10):1939 – 1942. <https://citeseerx.ist.psu.edu/viewdoc/download?doi=10.1.1.295.6387&rep=rep1&type=pdf>
26. Roshan Kumar Dubey, Hitesh Kumar Dewangan. Rational Design and Characterization of Transdermal Patch of Irbesartan for Hypertension. *Indian Journal of Pharmaceutical Education and Research*. 2020; 54(3): 464-472. <https://www.ijper.org/article/1259>
27. Priyanka Kriplani, Abhishek Sharma, Aman, Pooja Pun, Bhawna Chopra, AshwaniDhingra and GeetaDeswal. Formulation and Evaluation of Transdermal Patch of Diclofenac Sodium. *Global Journal of Pharmacy & Pharmaceutical Sciences*. 2018; 4(5): 01-04. DOI: 10.19080/GJPPS.2018.04.555647.
28. Mohd. Amjad, Mohd. Ehteshamuddin, S. Chand, Hanifa, M. Sabreesh, R. Asia, and G. S Kumar. Formulation and Evaluation of Transdermal Patches of Atenolol. *Advance Research in Pharmaceutics and Biologicals*. 2011; 1(2): 109-119. [https://arastirmax.com/en/system/files/dergiler/160852/makaleler/1/2/arastirmx\\_160852\\_1\\_pp\\_109-119.pdf](https://arastirmax.com/en/system/files/dergiler/160852/makaleler/1/2/arastirmx_160852_1_pp_109-119.pdf)
29. Hemul V. Patel, Jaimin D. Bhatt and Naynika K. Patel. Design and Development of Transdermal Drug Delivery for Anti-Hypertensive Drug Using Different Polymeric System. *International Journal of Pharmaceutical and Chemical Sciences*. 2013; 2(2):942-949. [https://www.researchgate.net/publication/265259719\\_Design\\_and\\_Development\\_of\\_Transdermal](https://www.researchgate.net/publication/265259719_Design_and_Development_of_Transdermal)

- Drug\_Delivery\_for\_Anti-Hypertensive\_Drug\_Using\_Different\_Polymeric\_System
30. TejashreeChavan and Prajakta More. Formulation and Evaluation of Transdermal patch of Antihypertensive Drug. International Journal of Science and Research. 2018; 7(7): 562-566.
31. Verma Surender, Malik Vipul, Ashima. Formulation, evaluation and optimization of transdermal patches of losartan potassium. World Journal of Pharmaceutical Sciences. 2016; 4(5): 277-284.[https://www.academia.edu/25197524/Formulation\\_evaluation\\_and\\_optimization\\_of\\_transdermal\\_patches\\_of\\_losartan\\_potassium](https://www.academia.edu/25197524/Formulation_evaluation_and_optimization_of_transdermal_patches_of_losartan_potassium)
32. VemulaVaishnavi, I. Bala Tripura Sundari and Dr. M. Bhagavan Raju. Formulation and Evaluation of Telmisartan Loaded Ethosomal Patch. European Journal of Pharmaceutical and Medical Research, 2019; 6(11): 407-416. [https://storage.googleapis.com/journal-uploads/ejpmr/article\\_issue/1572505805.pdf](https://storage.googleapis.com/journal-uploads/ejpmr/article_issue/1572505805.pdf)
33. Vaishali Y. Londhe and Kashmira B. Umalkar, Formulation Development and Evaluation of Fast Dissolving Film of Telmisartan, Indian Journal of Pharmaceutical Sciences, 2012, 74 (2), 122-126. doi: 10.4103/0250-474X.10384
34. S.C. Atram and Dr. S.D. Pande, A Statistical approach to the development of Telmisartan Transdermal Delivery System, Indo American Journal of Pharmaceutical Sciences, 2021, 08 (1), 1989-1998.<https://www.iajps.com/wp-content/uploads/2021/01/410.IAJPS410012021.pdf>
35. Swathi Palepu, Balli Sravanthi, Donthu Himabindu, GollapallyVenu Kumar, Jonnalagadda Rambabu, Dr T. Satyanarayana, Formulation and Evaluation of Pravastatin Sodium Transdermal Patch, Scholars Academic Journal of Pharmacy, 2017, 6(10), 440-445.<https://saspublishers.com/media/articles/SAJP-610440-445.pdf>
36. Rajesh Sreedharan Nair, Tai Nyet Ling, Mohamed Saleem Abdul Shukkoor, Balamurugan Manickam. Matrix type transdermal patches of captopril: Ex vivo permeation studies through excised rat skin. Journal of pharmacy research. 2013; 6: 774-779. <http://dx.doi.org/10.1016/j.jopr.2013.07.003>.
37. Shailesh T. Prajapati, Charmi G. Patel, and Chhagan N. Patel, Formulation and Evaluation of Transdermal Patch of Repaglinide, International Scholarly Research Network, 2011, 1-9. <https://downloads.hindawi.com/archive/2011/651909.pdf>.
38. M.D. Munoz, H. Castán, M.A. Ruiz, M.E. Morales, Design, development and characterization of transdermal patch of methadone, Journal of Drug Delivery Science and Technology. 2017; 42:1-6. <https://doi.org/10.1016/j.jddst.2017.04.011>.
39. Priyanka Arora, Biswajit Mukherjee, Design, Development, Physicochemical, and In Vitro and In Vivo Evaluation of Transdermal Patches Containing Diclofenac Diethylammonium Salt, Journal of Pharmaceutical Sciences, 2002, 91(9), 2076-2089. <https://doi.org/10.1002/jps.10200>.
40. Hany S.M. Ali, Ahmed F. Hanafy, Glibenclamide Nanocrystals in a Biodegradable Chitosan Patch for Transdermal Delivery: Engineering, Formulation, and Evaluation, Journal of Pharmaceutical Sciences, 2016, 402-410. <http://dx.doi.org/10.1016/j.xphs.2016.10.010>.
41. Dharmesh Trivedi, Anju Goyal. Formulation and evaluation of transdermal patches containing dexketoprofen Trometamol. International Journal of Pharmaceutical Chemistry and Analysis. 2020; 7(2):87-97. <https://doi.org/10.18231/j.ijpca.2020.014>.
42. S. Mutalik, N. Udupa, Glibenclamide Transdermal Patches: Physicochemical, Pharmacodynamic, and Pharmacokinetic Evaluations, Journal of Pharmaceutical Sciences, 2004, 93(6), 1577-1594. DOI: 10.1002/jps.20058.
43. Amandeep Singh and Alka Bali, Formulation and characterization of transdermal patches for controlled delivery of duloxetine hydrochloride, Journal of Analytical Science and Technology, 2016, 7(25), 1-13. DOI 10.1186/s40543-016-0105-6
44. Srinivas Mutalik, Nayanabhirama Udupa, Sharath Kumar, Sunil Agarwal, Ganesh Subramanian, Averineni K. Ranjith, Glipizide matrix transdermal systems for diabetes mellitus: Preparation, in vitro and preclinical studies, Science direct, 2006, 79, 1568-1577. <https://doi.org/10.1016/j.lfs.2006.05.002>.
45. Darren R. Gullick, W. John Pugh, Matthew J. Ingram, Paul A. Coxand Gary P. Moss. Formulation and characterization of a captopril ethyl ester drug-in-adhesive-type patch for percutaneous absorption. *Drug Development and Industrial Pharmacy*, 2010; 36(8): 926-932. DOI: 10.3109/03639040903585135.