

ALUMINIUM INDUCED OXIDATIVE STRESS AND ANTIOXIDANTS SYSTEM IN TWO BARLEY VARIETIES AND ITS ALLEVIATION THROUGH ASCORBIC ACID AND SALICYLIC ACID SEED PRIMING APPROACH

MD SHAHNAWAZ¹ AND DHEERA SANADHYA²*

¹ Ph.D. Res. Scholar, School of Life and Basic Sciences, Jaipur National University, Jaipur, Rajasthan, India ²* Associate Professor, School of Life and Basic Sciences, Jaipur National University, Jaipur, Rajasthan, India

ABSTRACT

The phytotoxic effect of various concentrations (2mM-6mM) of Al at acidic pH was investigated in terms of oxidative damage (ROS), molecules of lipid peroxidation (MDA, H₂O₂), non-enzymatic antioxidants (glutathione, flavonoids) and antioxidant enzyme activities (SOD, POD, Catalase) in two Barley varieties RD25522 (Al tolerant) and RD2052 (Al sensitive) and its alleviation was studied using seed priming approach with two PGR's Ascorbic acid (AA) and Salicylic acid (SA). Both AA and SA regulate the plant growth and protect the plant against biotic and abiotic stress due to their antioxidant activity. The experiment was carried out under *in vitro* conditions for six days. ROS content increased significantly with increased concentration of Aluminum. Biophysical parameters like Electrolyte leakage (EL) increased while Relative Leaf Water Content (RLWC) decreased due to increased ROS and also induced lipid peroxidation which was correlated with an increased membrane injury. The antioxidant enzymes like superoxide dismutase (SOD), ascorbate peroxidase (APX), catalase (CAT), and glutathion reductase (GR) showed increased activity while pyrogallol peroxidase (POD) and nonenzymatic antioxidant glutathione, flavonoids showed declined content with increased Al concentration. These result showed increased oxidative stress under induced Al phytotoxicity and barley variety RD2552 showed tolerant nature than RD2052 against Al stress.

Abbreviation- AA-Ascorbic Acid, SA-Salicylic Acid, SOD- Superoxide Dismutase, APX- Ascorbate Peroxidase, CAT- Catalase, GR- Glutathione Reductase, POD- Pyrogallol Peroxidase, RLWC- Relative Leaf Water Content, MDA- Malondialdehyde, H₂O₂-Hydrogen Peroxide, GSH- Glutathione

Keywords: Barley, Al toxicity, Ascorbic acid, Salicylic acid, Oxidative damage, Lipid peroxidation

INTRODUCTION

Aluminum (Al) is the third most abundant element in earth crust after oxygen and silicon approximately 8% of its mass. It is never present as free ion in soil though it is tightly bound with oxygen and silicon resulting in formation of nonphytotoxic and insoluble aluminum silicate and oxide forms. It is non essential and harmless in slightly acid or non acidic soil, because toxicity of Al is dependent upon soil acidity and soil acidity is one of the most important factors in soil that

constrains crop production in whole world. It is natural process that accelerates by agriculture practices. Soil acidity creates many problems for crop growth due to removal of crucial cations and minerals. It removes important cations Ca^{2+} , Mg^{2+} , K^+ , and Na^+ etc. and replace with H_3O and Al^{3+} in soil solution. The primary target of Al toxicity is root. It is reported that the even micromolar concentration of Al can inhibit root growth in many agriculturally important plant species. Active Al increases reactive oxygen species (ROS) production such as superoxide anion (O_2^-) , hydroxyl free

hydrogen peroxide (.OH), (H₂O₂).radical Excessive ROS generation causes disruption of lipid bilayer, nucleic acid damage and protein denaturation resulting in oxidative damage, physiological disorder and eventually premature senescence and cell death.⁵ It has been shown that Al exposure is associated with peroxidative damage of membrane lipids due to the stress-related increase in the production of highly toxic oxygen free radicals.⁶ Vitamin C or L-ascorbic acid, or simply ascorbate (the anion of ascorbic acid), is one of simplest and water soluble micronutrient essential for metabolic reactions in all animals and plants. Ascorbate has beneficial influences on various aspects in plants. It also participates in cell signaling modulation in numerous cellular processes including cell division, cell expansion and cell wall growth. Ascorbic acid neutralizes the excess ROS toxic effect by enzymatic as well as non enzymatic detoxification.⁸ Salicylic acid act as antioxidant and neutralize the harmful oxide radicals, and also, influences seed germination, cell growth, respiration, stomatal closure, senescenceassociated gene expression, responses to abiotic stresses, nodulation in legumes, and fruit yield.9 This study was based on effect of Al toxicity on two varieties of barley RD2552 and RD2052 analyzed the effect of two plant growth regulators (AA and SA) in mitigating the stress, and assess the biophysical (RLWC and EL), osmolyte (proline), signaling molecule (ROS), lipid peroxidation (MDA and H₂O₂), enzymatic (SOD, APX, CAT, POD and GR) and nonenzymatic antioxidants (GSH and Flavonoids) changes in plants.

MATERIAL AND METHODS

Study area and Plant material

The present work was carried out in Department of Life Sciences, Jaipur National University Jaipur, Rajasthan. Seeds of barley varieties (Hordeum VulgareL.) were collected from Rajasthan Agriculture Research Institute, Durgapura, Jaipur, Rajasthan. This study was targeted to analyze Al toxicity in two barley varieties (RD2052 and RD2552), primed with and without AA and SA. All seeds germinated in glass petri dishes with different concentrations of aluminum (C, 2mM, 4mM and 6mM) in ½ strength Hoagland solutions at pH4 for six days. After six days of germination analyzed the biophysical parameters, osmolyte (Proline), signaling molecule, lipid peroxidation, enzymatic and nonenzymatic antioxidant by following methods: RLWC, 10 EL, 11 Proline, 12 ROS, 13 MDA, 12

H₂O₂,¹⁵ Glutathione, ¹⁶ Flavonoids, ¹⁷ SOD, ¹⁸ APX, ¹⁹ POD, ²⁰ CAT, ²¹ GR. ²²

Data analysis

The data were determined by the one way analysis of variance (ANOVA), the design was completely randomized design (CRD). Data analysis was carried out using SPSS software. Vertical bar represent standard error.

RESULTS AND DISCUSSION

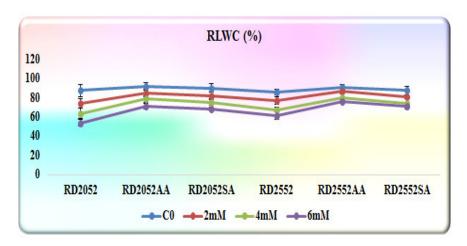
The one way analysis of variance (ANOVA) for all data determined that there were significant variation both varieties between (P<0.01). Aluminum toxicity decreased the RLWC, the effect increased with Al levels (Graph no.1). The same reduction has been reported in leaves of Sorghum and Mung bean,²³ resulting in loss of turgid and less water availability.²⁴ In susceptible variety RD2052 the reduction in the RLWC content at 6mM was 38.86% while in tolerant variety RD2552 it decreased by 29.16% over control. Priming with AA and SA improved RLWC (%) in both susceptible and the tolerant varieties over control compared with unprimed. Ekmekçi and Karaman, ²⁵ reported that AA treatment increased the RLWC content and reduction in transpiration rate indicating the efficiency of water uptake and reduction of water loss, which consequently causing increase in leaf water potential. Yalcin and Vardar,²⁴ reported that SA reduced Al toxicity and maintained reduced RWC content in Sorghum bicolor variety. Gondor et al., 26 reported in finger millet that supplementation of SA increased concentration of solutes in the cell causing improved water intake, high RWC and maintenance of osmotic balance. Aluminum toxicity increased the EL, the effect increased with Al level (Graph no.2). Surapu et al., ²⁷ also reported induced EL content and loss of membrane integrity in tomato cultivars. Tamas et al., 28 in barley showed that enhanced ROS generation caused increased electrolyte leakage and decreased cell viability. Similar results on EL were also reported in pea.²⁹ However, AA and SA significantly reduced membrane damage resulting in decreased EL compared to unprimed barley seedling under Al toxicity. Azooz, ³⁰ reported that ascorbic acid could accelerate cell division and cell enlargement and improved membrane integrity. Krantev et al., 31 reported that SA significantly reduced ion leakage by root and maintained cell membrane integrity in maize seedling. Proline content increased (Graph no.3) with increased

Aluminum concentration. Higher proline content was noticed in RD2052 than RD2552.Proline increase was reported to moderate aluminum by its osmolyte and antioxidant tension characteristic in Sunflower. 32 With AA and SA priming, proline content increased compared with unprimed. Amin et al., 33 reported that AA and SA increased the amount of proline in the plants leading to the resistance against water loss, protecting turgidity, reducing the membrane damage and accelerating growth of plants under stress conditions. Azzedine et al., 34 reported that application of ascorbic acid was effective to mitigate the adverse effect of salt stress on plant growth of durum wheat due to increased leaf area and enhanced proline accumulation. Misra and Saxena, 35 found that SA alleviated the salt stress of lentil by improving the proline metabolizing system. According to (Graph no.4) results obtained ROS content was increased with increasing Al concentration in both barley varieties. The production of ROS induced by Al is well-known phenomenon. ^{36, 37} Our results are in support with the findings that prove the subsequent accumulation of total ROS and lipid peroxides. Al is known to promote increased concentration of ROS and changes the redox state of the metabolic system in cells.³⁸ In experimental result AA and SA play crucial role in scavenging the ROS species. AA and SA showed reduction in ROS content in both varieties compared to unprimed variety. AA can be used by plants for the elimination of excess ROS produced during oxidative stress.³⁹ ROS species like O₂ and singlet oxygen can be eliminated through AA nonenzymatic defense mechanism. 40 Khan *et al.*, ⁴¹ reported that both endogenous and exogenous SA was evidenced to play roles in antioxidant metabolism and have a tight control over cellular ROS. The amount of MDA (Graph no.5) increased with the increase in Aluminum stress level, greater lipid peroxidation in RD2052 was observed than RD2552. Present results in agreement with those of Xiao et al., 42 who reported damage of the membrane system in Longan leaves under Al stress due to the induction of oxidative damage pertinent to the imbalance of ROS production. Sharma and Dubey, 43 reported that Al toxicity was associated with the Al-induced oxidative stress resultant increased generation of superoxide anion and H₂O₂ and enhanced MDA in rice shoot. Hosseini et al., 44 reported that Al toxicity increased the accumulation of lipid peroxidation product, MDA, which is regarded as an indicator of the loss of structural integrity in

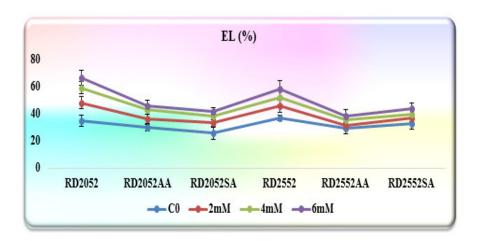
membranes subjected to heavy meal stress. Supplementation with AA and SA resulted in better recovery of seedling. Shao et al., 45 reported that Ascorbic acid protects metabolic processes against H₂O₂ and other toxic derivatives of oxygen affected many enzyme activities, minimize the damage caused by oxidative processes through synergistic function with other antioxidants and stabilize membranes. Salarizdah et al., 46 reported that SA led to a significant decrease in the level of lipid peroxidation. Al stress induced a significant increase in hydrogen peroxide (H₂O₂) level in both barley seedlings. According to (Graph no.6) the experimental result in barley seedling under Al toxicity, The RD2052 variety showed significantly increased H₂O₂ content than RD2552 variety with aluminum treatment at 6mM concentration compared to control, but AA and SA treated value was less increased in both varieties compared to unprimed control at 6mM Aluminum. Ascorbic acid can directly scavenge superoxide, hydroxyl radicals and singlet oxygen and reduced H₂O₂ to water via ascorbate peroxidase reaction, and is the major antioxidant that scavenges H₂O₂ produced by ROS and plays important roles in plants under abiotic stress tolerance. 47 Belkhadi et al., 48 reported that an increased tolerance of Flax to Cd was attributed to SA-mediated control of H₂O₂ accumulation. GSH level is an important protective mechanism to minimize oxidative damage in plants exposed to metals. It plays a vital role in the antioxidant defense system as well as the glyoxalase system by acting as a substrate or cofactor for certain enzymes. A marked decrease in GSH content with increased Al concentration of the Barley (RD2052 and RD2552) shoot and root was observed under Al stress (6mM) when compared to the controls (Graph no.7). The results are in agreement with Noctor et al., 49 and Ezaki et al., 50. Glutathione content significantly maintained in AA, SA treated Shoot and Root of RD 2052 and RD2552 at 6mM Al concentration compared to unprimed barley seedlings. Ascorbic acid and glutathione both are antioxidants that increase in the activity of antioxidant enzymes and scavenging the ROS species, The AA-GSH cycle is a mechanism for removing H₂O₂. 51 Similar observations of SA mediated amelioration of drought stress were reported in mustard and mung bean by improved antioxidant system. 52 SA-application (at 0.5mM) increased the activity of enzymes of AsA-GSH pathway resulted in the increased tolerance of B. *juncea* to salinity stress. 53 Flavonids are one class of secondary metabolites that are also known as

Vitamin P. In our result Flavonoids content (Graph no.8) was also decreased with increased Aluminum concentration, similar result has been found by Zahra et al., 54. Flavonoid was significantly decreased in both verities RD2052 and RD2552. In the same way AA and SA primed seedlings showed maintained flavonoid content compared to unprimed barley seedling at 6mM Al. Major ROSscavenging enzymes in seedlings include SOD, APX and CAT. 55, 56 The most effective antioxidative enzyme in preventing ROS induced cellular oxidative damage is Superoxide dismutase (SOD) which catalyzes the conversion of O_2^- to H₂O₂, whereas CAT scavenges H₂O₂ and peroxidase uses H₂O₂ for the oxidation of various inorganic and organic substrates.⁵⁷ The present studies (Graph no. 9-13) demonstrated significant increase in Catalase, Ascorbate Peroxidase, SOD, Glutathione Reductase activity, while decreased Pyrogallol Peroxidase activities in barley seedlings with increased Aluminum concentration. Increased SOD, Catalase, Ascorbate Peroxidase, Glutathione Reductase activity in RD2052 was more than RD2552, while Pyrogallol Peroxidase activity decreased in RD2052 more than RD2552. Similarly

in maize and wheat roots Al treatment also led to the increase of ROS production and simultaneously increased SOD, CAT and APX. 57, 58 Batool et al., 5 reported that decreased the activity of POD under salt stress. Divya et al., 60 reported decreased POD activity with Al toxicity. Bor et al., 61 reported that increased activity of GR is closely related with salt capacity in sugar beet tolerance Supplementation with AA and SA, the enzyme activity gradually changed due to their ameliorative effect. SOD activity gradually decreased on supplement with AA and SA, indicating lower ROS accumulation to scavenge. AA and SA maintain the CAT activity in Al exposed seedling, there by facilitating dismutation of H₂O₂ into non toxic form so as to minimize the ROS induced oxidative stressed. AA acting as a nonenzymatic antioxidant, scavenges the ROS species, and enzymes requires AA as a cofactor in the detoxifying enzymatic processes. 62 AA can directly scavenge superoxide. hydroxyl radicals and singlet oxygen and diminish H₂O₂ to water via ascorbate peroxidase reaction.⁶² Hayat et al., 63 reported that SA enhanced the activities of antioxidative enzymes and protected the plants against oxidative damage.



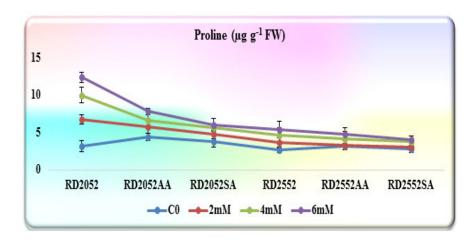
Graph no.1
RLWC (%) in RD2052 and RD2552 Barley varieties (unprimed and primed with AA and SA)
germinated under different concentrations of Aluminum at pH₄



Graph no.2

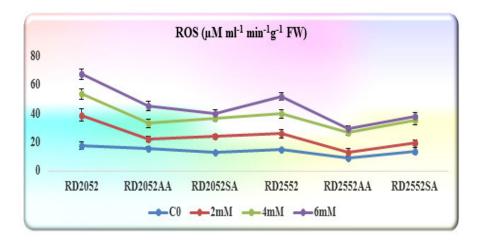
EL (%) in RD2052 and RD2552 Barley varieties (unprimed and primed with AA and SA)

germinated under different concentrations of Aluminum at pH₄

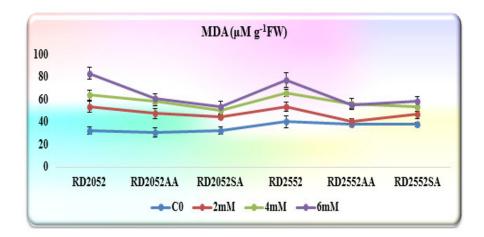


Graph no.3

Proline content ($\mu g \ g^{-1}FW$) in RD2052 and RD2552 Barley varieties (unprimed and primed with AA and SA) germinated under different concentrations of Aluminum at pH_4

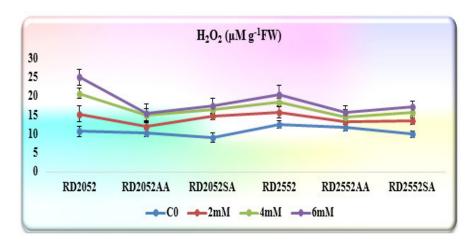


Graph no.4 ROS content ($\mu M \ ml^{-1} \ min^{-1} \ g^{-1} \ FW$) in RD2052 and RD2552 Barley varieties (unprimed and primed with AA and SA) germinated under different concentrations of Aluminum at pH₄

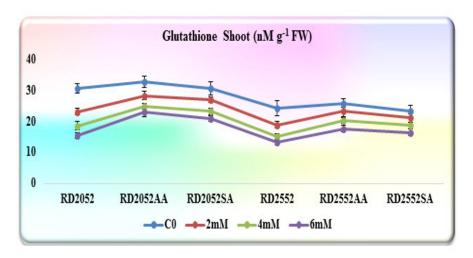


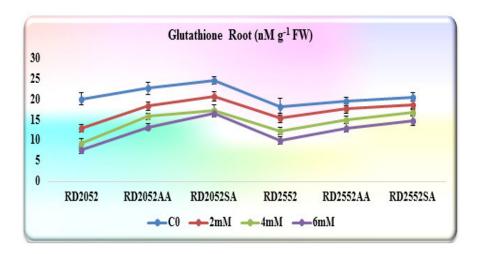
Graph no.5

MDA content (μ M $g^{-1}FW$) in RD2052 and RD2552 Barley varieties (unprimed and primed with AA and SA) germinated under different concentrations of Aluminum at pH_4



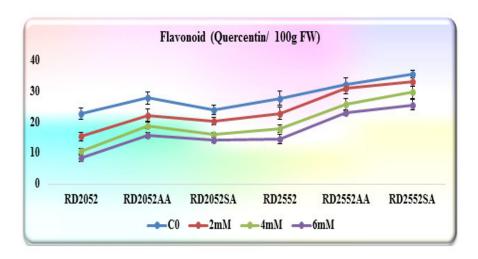
Graph no.6 H_2O_2 content (μM $g^{-1}FW$) in RD2052 and RD2552 Barley varieties (unprimed and primed with AA and SA) germinated under different concentrations of Aluminum at pH_4



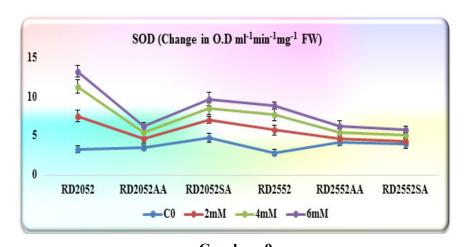


Graph no.7

Glutathione content (nM $g^{-1}FW$) in RD2052 and RD2552 Barley varieties (unprimed and primed with AA and SA) germinated under different concentrations of Aluminum at pH₄

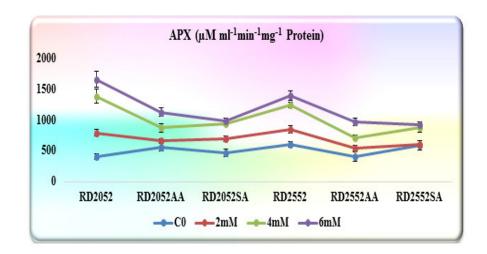


Graph no.8 Flavonoid (Quercentin/ 100g FW) in RD2052 and RD2552 Barley varieties (unprimed and primed with AA and SA) germinated under different concentrations of Aluminum at pH $_4$

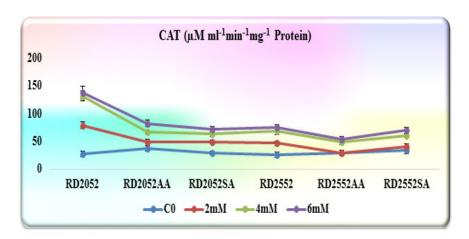


Graph no.9

SOD activity (Change in O.D ml⁻¹min⁻¹mg⁻¹ FW) in RD2052 and RD2552 Barley varieties (unprimed and primed with AA and SA) germinated under different concentrations of Aluminum at pH₄

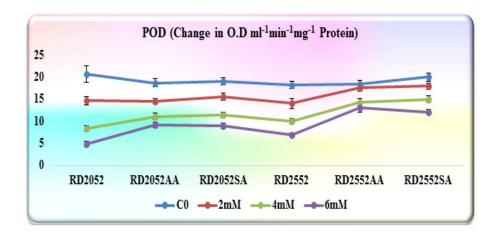


Graph no.10 APX activity (μM m Γ^{1} min $^{-1}$ mg $^{-1}$ Protein) in RD2052 and RD2552 Barley varieties (unprimed and primed with AA and SA) germinated under different concentrations of Aluminum at pH₄



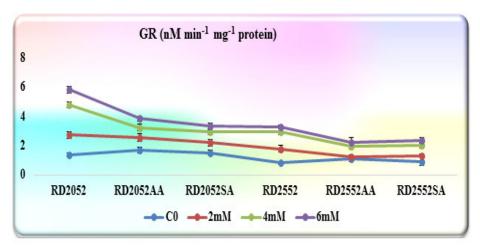
Graph no.11

CAT activity (µM ml¹min⁻¹mg⁻¹ Protein) in RD2052 and RD2552 Barley varieties (unprimed and primed with AA and SA) germinated under different concentrations of Aluminum at pH₄



Graph no.12

POD (Change in O.D mt⁻¹min⁻¹mg⁻¹ Protein) in RD2052 and RD2552 Barley varieties (unprimed and primed with AA and SA) germinated under different concentrations of Aluminum at pH₄



Graph no.13

GR activity (nM min⁻¹ mg⁻¹ protein) in RD2052 and RD2552 Barley varieties (unprimed and primed with AA and SA) germinated under different concentrations of Aluminum at pH₄

CONCLUSION

The result of the experiment reveled that AA and SA seed priming successfully mitigated the Al toxicity in both barley varieties (RD2052 and RD2552). The antioxidative property of both the

PGRs minimized the oxidative damage due to Al toxicity. Seed priming is a convenient and economical approach that can be further used for field experiments. RD2552 proved its tolerant nature by sustaining the harmful effect of Al and performed better than RD2052 against Al stress.

REFERENCES

- 1. Verstraeten SV, Aimo L, Oteiza PI. Aluminium and lead: molecular mechanisms of brain toxicity. Archives of toxicology. 2008 Nov 1;82(11):789-802.
- 2. Sun P, Tian QY, Chen J, Zhang WH. Aluminium-induced inhibition of root elongation in Arabidopsis is mediated by ethylene and auxin. Journal of Experimental Botany. 2010;61(2):347-56.
- 3. Jovanovic Ž, Djalovic I, Komljenovic I, Kovacevic V, Cvijović M. Influences of liming on vertisol properties and yields of the field crops. Cereal Research Communications. 2006 Mar 1;34(1):517-20.
- 4. Jovanovic Z, Tolimir M, Đalovic I, Cvijovic M. Influences of growing system and NPK-fertilization on maize yield on pseudogley of central Serbia. Cereal Research Communications. 2007 Apr 25;35(2):1329-32.
- 5. Wang ZH, Liang QL, Wang YM, Luo GA. Carbon nanotube-intercalated graphite electrodes for simultaneous determination of dopamine and serotonin in the presence of ascorbic acid. Journal of Electroanalytical Chemistry. 2003 Jan 2;540:129-34.

- 6. Apel K, Hirt H. Reactive oxygen species: metabolism, oxidative stress, and signal transduction. Annu. Rev. Plant Biol.. 2004 Jun 2:55:373-99.
- 7. Qian W, Yu C, Qin H, Liu X, Zhang A, Johansen IE, Wang D. Molecular and functional analysis of phosphomannomutase (PMM) from higher plants and genetic evidence for the involvement of PMM in ascorbic acid biosynthesis in Arabidopsis and Nicotiana benthamiana. The Plant Journal. 2007 Feb 1;49(3):399-413.
- 8. Mittler R. Oxidative stress, antioxidants and stress tolerance. Trends in plant science. 2002 Sep 1;7(9):405-10.
- 9. Hayat Q, Hayat S, Irfan M, Ahmad A. Effect of exogenous salicylic acid under changing environment: a review. Environmental and experimental botany. 2010 Mar 31;68(1):14-25.
- 10. WEATHERLEY P. Studies in the water relations of the cotton plant. New Phytologist. 1950 Mar 1;49(1):81-97.
- 11. Lin CY, Chen YM, Key JL. Solute leakage in soybean seedlings under various heat shock

- regimes. Plant and cell physiology. 1985 Dec 1;26(8):1493-8.
- 12. Bates LS, Waldren RP, Teare ID. Rapid determination of free proline for water-stress studies. Plant and soil. 1973 Aug 1;39(1):205-7.
- 13. Able AJ, Guest DI, Sutherland MW. Use of a New Tetrazolium-Based Assay to Study the Production of Superoxide Radicals by Tobacco Cell Cultures Challenged with Avirulent Zoospores of Phytophthora parasitica varnicotianae. Plant Physiology. 1998 Jun 1;117(2):491-9.
- 14. Heath RL, Packer L. Photoperoxidation in isolated chloroplasts: I. Kinetics and stoichiometry of fatty acid peroxidation. Archives of biochemistry and biophysics. 1968 Apr 1;125(1):189-98.
- 15. Mukherjee SP, Choudhuri MA. Implications of water stress-induced changes in the levels of endogenous ascorbic acid and hydrogen peroxide in Vigna seedlings. Physiologia Plantarum. 1983 Jun 1;58(2):166-70.
- 16. Anderson ME. [70] Determination of glutathione and glutathione disulfide in biological samples. Methods in enzymology. 1985 Dec 31;113:548-55.
- 17. Marinova D, Ribarova F, Atanassova M. Total phenolics and total flavonoids in Bulgarian fruits and vegetables. Journal of the university of chemical technology and metallurgy. 2005 Jul;40(3):255-60.
- 18. DHINDSA RS, Plumb-Dhindsa P, THORPE TA. Leaf senescence: correlated with increased levels of membrane permeability and lipid peroxidation, and decreased levels of superoxide dismutase and catalase. Journal of Experimental botany. 1981 Feb 1;32(1):93-101..
- 19. Nakano Y, Asada K. Hydrogen peroxide is scavenged by ascorbate-specific peroxidase in spinach chloroplasts. Plant and cell physiology. 1981 Aug 1;22(5):867-80.
- 20. Chance B, Maehly AC. [136] Assay of catalases and peroxidases. Methods in enzymology. 1955 Dec 31;2:764-75.
- 21. Aebi H. [13] Catalase in vitro. Methods in enzymology. 1984 Dec 31;105:121-6.
- 22. Carlberg I, Mannervik B. [59] Glutathione reductase. Methods in enzymology. 1985 Dec 31;113:484-90.
- 23. Ali B, Hasan SA, Hayat S, Hayat Q, Yadav S, Fariduddin Q, Ahmad A. A role for brassinosteroids in the amelioration of

- aluminium stress through antioxidant system in mung bean (Vigna radiata L. Wilczek). Environmental and Experimental Botany. 2008 Mar 31;62(2):153-9.
- 24. Yalcin G, Vardar F. The alleviating effects of salicylic acid application against aluminium toxicity in barley (Hordeum vulgare) roots. Biologia. 2016 Dec 1;71(12):1338-44.
- 25. Ekmekçi BA, Karaman M. Exogenous ascorbic acid increases resistance to salt of Silybum marianum (L.). African Journal of Biotechnology. 2012;11(42):9932-40.
- 26. Gondor OK, Pál M, Darko E, Janda T, Szalai G. Salicylic Acid and Sodium Salicylate Alleviate Cadmium Toxicity to Different Extents in Maize (Zea mays L.). PloS one. 2016 Aug 4;11(8):e0160157.
- 27. Surapu V, Ediga A, Meriga B. Salicylic acid alleviates aluminum toxicity in tomato seedlings (Lycopersicum esculentum Mill.) through activation of antioxidant defense system and proline biosynthesis. Advances in Bioscience and Biotechnology. 2014 Aug 8;2014.
- 28. Tamas L, Budikova S, Simonovicova M, Huttova J, Siroka B, Mistrik I. Rapid and simple method for Al-toxicity analysis in emerging barley roots during germination. Biologia plantarum. 2006 Mar 1;50(1):87-93.
- 29. Yamamoto Y, Kobayashi Y, Matsumoto H. Lipid peroxidation is an early symptom triggered by aluminum, but not the primary cause of elongation inhibition in pea roots. Plant Physiology. 2001 Jan 1;125(1):199-208.
- 30. Azooz MM, Al-Fredan MA. The inductive role of vitamin C and its mode of application on growth, water status, antioxidant enzyme activities and protein patterns of Vicia faba L cv. Hassawi grown under seawater irrigation. Amer J Plant Physiol. 2009;4:38-51.
- 31. Krantev A, Yordanova R, Janda T, Szalai G, Popova L. Treatment with salicylic acid decreases the effect of cadmium on photosynthesis in maize plants. Journal of plant physiology. 2008 Jun 16;165(9):920-31.
- 32. Ziaei N, Rezaiatmand Z, Ranjbar M. Study of Aluminum Toxicity on Photosynthetic Pigment, Soluble Sugars and Proline Contents in Two Sunflower Varieties. Res. Crop Ecophysiol.. 2014;9(2):105-13.
- 33. Amin B, Mahleghah G, Mahmood HM, Hossein M. Evaluation of interaction effect of drought stress with ascorbate and salicylic acid on some of physiological and

- biochemical parameters in okra (Hibiscus esculentus L.). Res. J. Biol. Sci. 2009;4:380-7.
- 34. Azzedine F, Gherroucha H, Baka M. Improvement of salt tolerance in durum wheat by ascorbic acid application. Journal of stress physiology & Biochemistry. 2011;7(1).
- 35. Misra N, Saxena P. Effect of salicylic acid on proline metabolism in lentil grown under salinity stress. Plant Science. 2009 Sep 30;177(3):181-9.
- 36. Xu FJ, Li G, Jin CW, Liu WJ, Zhang SS, Zhang YS, Lin XY. Aluminum-induced changes in reactive oxygen species accumulation, lipid peroxidation and antioxidant capacity in wheat root tips. Biologia plantarum. 2012 Mar 1;56(1):89-96.
- 37. Yamamoto Y, Kobayashi Y, Devi SR, Rikiishi S, Matsumoto H. Aluminum toxicity is associated with mitochondrial dysfunction and the production of reactive oxygen species in plant cells. Plant physiology. 2002 Jan 1;128(1):63-72.
- 38. Ma B, Gao L, Zhang H, Cui J, Shen Z. Aluminum-induced oxidative stress and changes in antioxidant defenses in the roots of rice varieties differing in Al tolerance. Plant cell reports. 2012 Apr 1;31(4):687-96.
- Gratão PL, Polle A, Lea PJ, Azevedo RA. Making the life of heavy metal-stressed plants a little easier. Functional Plant Biology. 2005 Jul 8;32(6):481-94.
- 40. Devi SR, Yamamoto Y, Matsumoto H. An intracellular mechanism of aluminum tolerance associated with high antioxidant status in cultured tobacco cells. Journal of Inorganic Biochemistry. 2003 Sep 15;97(1):59-68.
- 41. Khan MI, Asgher M, Khan NA. Alleviation of salt-induced photosynthesis and growth inhibition by salicylic acid involves glycinebetaine and ethylene in mungbean (Vigna radiata L.). Plant Physiology and Biochemistry. 2014 Jul 31;80:67-74.
- 42. Xiao X, Liu X, Yang Z, Chen L, Cai Y. Effect of aluminum stress on the photosynthesis of longan seedlings. Chinese Journal of Tropical Crops. 2004 Dec 26(1):63-9.
- 43. Hosseini M, Boojar MM, Delkhosh B, Moradi P. Effects of foliar spraying acetyl-coA on dry weight of leaves, chlorophyll a, and antioxidant enzymes of rosemary

- (Rosmarinus officinalis L.). Journal of Plant Physiology. 2014 Jan 1;4(2):977-81.
- 44. Shao HB, Chu LY, Lu ZH, Kang CM. Primary antioxidant free radical scavenging and redox signaling pathways in higher plant cells. Int J Biol Sci. 2008 Jan 1;4(1):8-14.
- 45. Salarizdah M, Baghizadeh A, Abasi F, Mozaferi H, Salarizdah S. Response of Brassica napus L grains to the interactive effect of salinity and salicylic acid. Journal of Stress Physiology & Biochemistry. 2012;8(2).
- 46. Beltagi MS. Exogenous ascorbic acid (vitamin C) induced anabolic changes for salt tolerance in chick pea (Cicer arietinum L.) plants. African Journal of Plant Science. 2008 Oct 31;2(10):118-23.
- 47. Belkhadi A, Hediji H, Abbes Z, Nouairi I, Barhoumi Z, Zarrouk M, Chaïbi W, Djebali W. Effects of exogenous salicylic acid pretreatment on cadmium toxicity and leaf lipid content in Linum usitatissimum L. Ecotoxicology and environmental safety. 2010 Jul 31;73(5):1004-11.
- 48. Ezaki B, Suzuki M, Motoda H, Kawamura M, Nakashima S, Matsumoto H. Mechanism of gene expression of Arabidopsis glutathione Stransferase, AtGST1, and AtGST11 in response to aluminum stress. Plant physiology. 2004 Apr 1;134(4):1672-82.
- 49. Noctor G, Arisi AC, Jouanin L, Kunert KJ, Rennenberg H, Foyer CH. Glutathione: biosynthesis, metabolism and relationship to stress tolerance explored in transformed plants. Journal of Experimental Botany. 1998 Apr 1;49(321):623-47.
- 50. Foyer CH, Noctor G. Ascorbate and glutathione: the heart of the redox hub. Plant physiology. 2011;155(1):2-18.
- 51. Liu J, Piñeros MA, Kochian LV. The role of aluminum sensing and signaling in plant aluminum resistance. Journal of Integrative Plant Biology. 2014 Mar 1;56(3):221-30.
- 52. Nazar R, Umar S, Khan NA. Exogenous salicylic acid improves photosynthesis and growth through increase in ascorbate-glutathione metabolism and S assimilation in mustard under salt stress. Plant signaling & behavior. 2015 Mar 4;10(3):e1003751.
- 53. Zahra T, Amin B, Shekofeh E. The Effects of Aluminum and Phosphorous on some of Physiological Characteristics of Brassica napus. Journal of Stress Physiology & Biochemistry. 2015;11(1).

- 54. Willekens H, Chamnongpol S, Davey M, Schraudner M, Langebartels C, Van Montagu M, Inzé D, Van Camp W. Catalase is a sink for H2O2 and is indispensable for stress defence in C3 plants. The EMBO journal. 1997 Aug 15;16(16):4806-16.
- 55. Anjum NA, Umar S, Chan MT, editors. Ascorbate-glutathione pathway and stress tolerance in plants. Springer Science & Business Media; 2010 Aug 20.
- 56. Babourina O, Ozturk L, Cakmak I, Rengel Z. Reactive oxygen species production in wheat roots is not linked with changes in H+ fluxes during acidic and aluminium stresses. Plant signaling & behavior. 2006 Mar 1;1(2):70-5.
- 57. Boscolo PR, Menossi M, Jorge RA. Aluminum-induced oxidative stress in maize. Phytochemistry. 2003 Jan 31;62(2):181-9.
- 58. Ejaz B, Sajid ZA, Aftab F. Effect of exogenous application of ascorbic acid on antioxidant enzyme activities, proline contents, and growth parameters of Saccharum spp. hybrid cv. HSF-240 under salt stress. Turkish Journal of Biology. 2012 Nov 21;36(6):630-40.
- 59. Sri ND, Mohan MM, Mahesh K, Raghu K, Rao SS. Amelioration of Aluminium Toxicity

- in Pigeon Pea [Cajanus cajan (L.) Millsp.] Plant by 24-Epibrassinolide. American Journal of Plant Sciences. 2016 Aug 19:7(12):1618.
- 60. Conklin PL, Barth C. Ascorbic acid, a familiar small molecule intertwined in the response of plants to ozone, pathogens, and the onset of senescence. Plant, Cell & Environment. 2004 Aug 1;27(8):959-70.
- 61. Sen P, Aich A, Pal A, Sen S, Pal D. Profile of Antioxidants and Scavenger Enzymes during Different Developmental Stages in Vigna radiata (L.) Wilczek (Mungbean) under Natural Environmental Conditions. International Journal of Plant Research. 2014;4(2):56-61.
- 62. Khan MI, Asgher M, Khan NA. Alleviation of salt-induced photosynthesis and growth inhibition by salicylic acid involves glycinebetaine and ethylene in mungbean (Vigna radiata L.). Plant Physiology and Biochemistry. 2014 Jul 31;80:67-74.
- 63. Hayat S, Hasan SA, Fariduddin Q, Ahmad A. Growth of tomato (Lycopersicon esculentum) in response to salicylic acid under water stress. Journal of Plant Interactions. 2008 Dec 1;3(4):297-304.